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(54) Title: SYNTHETIC MAMMALIAN α-N-ACETYLO	GLUCC	AMINIDASE AND GENETIC SEQUENCES ENCODING	SAME

(57) Abstract

The present invention relates generally to mammalian α -N-acetyglucosaminidase and to genetic sequences encoding same and to their use in the investigation, diagnosis and treatment of subjects suspected of or suffering from α -N-acetyglucosaminidase deficiency.

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WO 97/19177 PCT/AU96/00747

SYNTHETIC MAMMALIAN α-N-ACETYLGLUCOSAMINIDASE AND GENETIC SEQUENCES ENCODING SAME

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FIELD OF THE INVENTION

The present invention relates generally to mammalian α -N-acetylglucosaminidase and to genetic sequences encoding same and to the use of these in the investigation, diagnosis and treatment of subjects suspected of or suffering from α -N-acetylglucosaminidase deficiency.

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Bibliographic details of the publications referred to by author in this specification are collected at the end of the description. Sequence Identity Numbers (SEQ ID NOs.) for the nucleotide and amino acid sequences referred to in the specification are defined following the bibliography.

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Throughout this specification, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated element or integer or group of elements or integers but not the exclusion of any other element or integer or group of elements or integers.

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BACKGROUND TO THE INVENTION

The increasing sophistication of recombinant DNA technology is greatly facilitating the efficacy of many commercially important industries including areas of medical and pharmaceutical research and development. The ability to purify native proteins and subsequently clone genetic sequences encoding these proteins is an important first step in the development of a range of therapeutic and diagnostic procedures. However, practitioners have faced many difficulties in purifying target molecules to an extent sufficient to determine amino acid sequences to permit the development of oligonucleotide probes to assist in the cloning of genetic sequences encoding the target molecules.

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Such difficulties have been particularly faced in the research and development of lysosomal enzymes. An important lysosomal enzyme is α -N-acetylglucosaminidase (EC 2.1.50). This enzyme acts as a exoglycosidase in lysosomes to hydrolyse the terminal α -N-acetylglucosamine residues present at the non-reducing terminus of fragments of heparan sulphate and heparin (Hopwood, 1989). A deficiency in this lysosomal hydrolase is responsible for the pathogenesis of Sanfilippo B (Mucopolysaccharidosis type IIIB [MPS-IIIB]) syndrome (von-Figura and Kresse, 1972; O'Brien, 1972). This is an autosomal recessive disorder of glycosaminoglycan catabolism leading to storage and excretion of excessive amounts of heparan sulphate and a variety of clinical phenotypes, but classically presenting with progressive mental retardation in conjunction with skeletal deformities (McKusick and Neufeld, 1983).

There is a need, therefore, to purify α -N-acetylglucosaminidase and to clone genetic sequences encoding same to permit development of a range of therapeutic and diagnostic procedures to assist in the diagnosis and treatment of disease conditions arising from α -N-acetylglucosaminidase deficiency.

20 SUMMARY OF THE INVENTION

One aspect of the invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides which encodes or is complementary to a sequence which encodes a mammalian α -N-acetylglucosaminidase or fragment or derivative thereof.

- A second aspect of the invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides which is capable of hybridising under at least low stringency conditions to a nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID NO:3 or a complementary strand or a homologue, analogue or derivative thereof.
- Another aspect of the invention is directed an isolated nucleic acid molecule which is at least 40% identical to the nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID

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NO:3 or to a complementary strand thereof or a homologue, analogue or derivative thereof.

A further aspect of the present invention provides a nucleic acid molecule comprising a sequence of nucleotides encoding or complementary to a sequence encoding a polypeptide capable of hydrolysing the terminal α -N-acetylglucosamine residues present at the non-reducing terminus of fragments of heparan sulphate and heparin residues and wherein said nucleotide sequence is capable of hybridising under low stringency conditions to the nucleotide sequence set forth in SEQ ID NO:1.

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A further aspect of the invention is directed to a genetic construct comprising a sense molecule, for the expression or over-expression of α -N-acetylglucosaminidase in prokaryotic or eukaryotic cells.

15 A further aspect of the present invention is directed to synthetic α-N-acetylglucosaminidase or like molecule.

A further aspect of the invention contemplates antibodies to α -N-acetylglucosaminidase and preferably synthetic α -N-acetylglucosaminidase or a like molecule.

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In still yet another aspect of the present invention there is contemplated a method of diagnosing a mutation or other abberations in the α -N-acetylglucosaminidase gene in a human or animal patient.

- 25 Another aspect contemplates a method of treating patients suffering from α-N-acetylglucosaminidase deficiency, such as in MPS-IIIB, said method comprising administering to said patient an effective amount of α-N-acetylglucosaminidase or active like form thereof.
- 30 Another aspect of the present invention is directed to a pharmaceutical composition comprising a recombinant mammalian α-N-acetylglucosaminidase or an active fragment

or derivative thereof and one or more pharmaceutically acceptable carriers and/or diluents.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a photographic representation of α -N-acetylglucosaminidase purified from human placenta following SDS/PAGE. Lane 1: M, standards (kDa); Lanes 2 and 3: purified α -N-acetylglucosaminidase from human placenta. Lane 4 and 5, bovine serum albumin.

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Figure 2 is a photographic representation of an SDS/polyacrylamide gel showing the molecular weights of recombinant α -N-acetylglucosaminidase polypeptides produced in CHO cells before (-) and after (+) PNGase F digestion. The 50 mM NaCl and 75 mM NaCl fractions are indicated. Molecular weights of α -N-acetylglucosaminidase polypeptides are indicated on the left of the figure. Molecular weights of marker proteins are indicated on the right hand side of the figure (lane 5).

Single and three letter abbreviations of conventional amino acid residues as used herein are defined in Table 1.

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Suitable amino acid substitutions referred to herein are defined in Table 2.

Codes for non-conventional amino acid residues as used herein are defined in Table 3.

- 5 -

TABLE 1

Amino Acid	Three-letter Abbreviation	One-letter Symbol
Alanine	Ala	Α
Arginine	Arg	R
Asparagine	Asn	N
Aspartic acid	Asp	D
Cysteine	Cys	C
Glutamine	Gln	Q
Glutamic acid	Glu	Е
Glycine	Gly	G
Histidine	His	Н
Isoleucine	Ile	I
Leucine	Leu	· L
Lysine	Lys	K
Methionine	Met	M
Phenylalanine	Phe	F
Proline	Pro	P
Serine	Ser	S
Threonine	Thr	T
Tryptophan	Тгр	w
Tyrosine	Туг	Y
Valine	Val	v
Any residue	Xaa	X

WO 97/19177 PCT/AU96/00747

- 6 -

TABLE 2
Suitable residues for amino acid substitutions

	Original Residue	Exemplary Substitutions
5	Ala	Ser
	Arg	Lys
	Asn	Gln; His
	Asp	Glu
	Cys	Ser
10	Gln	Asn
	Glu	Asp
	Gly	Pro
	His	Asn, Gln
	Ile	Leu; Val
15	Leu	Ile; Val
	Lys	Arg; Gln; Glu
	Met	Leu; Ile
	Phe	Met; Leu; Tyr
•	Ser	Thr
20	Thr	Ser
	Trp	Tyr
	Tyr	Trp; Phe
	Val	Ile; Leu

-7-

TABLE 3

Non-conventional amino acid	Code	Non-conventional amino acid	Code
α-aminobutyric acid	Abu	L-N-methylalanine	Nmala
α -amino- α -methylbutyrate	Mgabu	L-N-methylarginine	Nmarg
aminocyclopropane-	Cpro	L-N-methylasparagine	Nmasn
carboxylate	* ** * *	L-N-methylaspartic acid	Nmasp
aminoisobutyric acid	Aib	L-N-methylcysteine	Nmcys
aminonorbornyl-	Norb	L-N-methylglutamine	Nmgin
carboxylate		L-N-methylglutamic acid	Nmglu
cyclohexylalanine	Chexa	L-N-methylhistidine	Nmhis
cyclopentylalanine	Cpen	L-N-methylisolleucine	Nmile
D-alanine	Dal	L-N-methylleucine	Nmleu
D-arginine	Darg	L-N-methyllysine	Nmlys
D-aspartic acid	Dasp	L-N-methylmethionine	Nmmet
D-cysteine	Dcys	L-N-methylnorleucine	Nmnle
D-glutamine	Dgln	L-N-methylnorvaline	Nmnva
D-glutamic acid	Dglu	L-N-methylornithine	Nmorn
D-histidine	Dhis	L-N-methylphenylalanine	Nmphe
D-isoleucine	Dile	L-N-methylproline	Nmpro
D-leucine	Dleu	L-N-methylserine	Nmser
D-lysine	Dlys	L-N-methylthreonine	Nmthr
D-methionine	Dmet	L-N-methyltryptophan	Nmtrp
D-ornithine	Dorn	L-N-methyltyrosine	Nmtyr
D-phenylalanine	Dphe	L-N-methylvaline	Nmval
D-proline	Dpro	L-N-methylethylglycine	Nmetg
D-serine	Dser	L-N-methyl-t-butylglycine	Nmtbug
D-threonine	Dthr	L-norleucine	Nle
D-tryptophan	Dtrp	L-norvaline	Nva
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	D-tyrosine	Dtyr	α-methyl-aminoisobutyrate	Maib
	D-valine	Dval	α-methyl-γ-aminobutyrate	Mgabu
	D-α-methylalanine	Dmala	α-methylcyclohexylalanine	Mchexa
	D-α-methylarginine	Dmarg	α-methylcylcopentylalanine	Mcpen
5	$D-\alpha$ -methylasparagine	Dmasn	α-methyl-α-napthylalanine	Manap
	D-α-methylaspartate	Dmasp	α-methylpenicillamine	Mpen
	D-α-methylcysteine	Dmcys	N-(4-aminobutyl)glycine	Nglu
	D-α-methylglutamine	Dmgln	N-(2-aminoethyl)glycine	Naeg
	D-α-methylhistidine	Dmhis 🚉	N-(3-aminopropyl)glycine	Norn
10	D-α-methylisoleucine	Dmile	N-amino-α-methylbutyrate	Ňmaabu
	D-α-methylleucine	Dmleu	α-napthylalanine	Anap
	D-α-methyllysine	Dmlys	N-benzylglycine	Nphe
	D-\a-methylmethionine	Dmmet	N-(2-carbamylethyl)glycine	Ngln
	D-\alpha-methylornithine	Dmorn	N-(carbamylmethyl)glycine	Nasn
15	D-α-methylphenylalanine	Dmphe	N-(2-carboxyethyl)glycine	Nglu
	D-α-methylproline	Dmpro	N-(carboxymethyl)glycine	Nasp
	D-α-methylserine	Dmser	N-cyclobutylglycine	Ncbut
	D-α-methylthreonine	Dmthr	N-cycloheptylglycine	Nchep
	D-α-methyltryptophan	Dmtrp	N-cyclohexylglycine	Nchex
20	D-α-methyltyrosine	Dmty	N-cyclodecylglycine	Ncdec
	D-α-methylvaline	Dmval	N-cylcododecylglycine	Ncdod
	D-N-methylalanine	Dnmala	N-cyclooctylglycine	Ncoct
	D-N-methylarginine	Dnmarg	N-cyclopropylglycine	Ncpro
	D-N-methylasparagine	Dnmasn	N-cycloundecylglycine	Nound
25	D-N-methylaspartate	Dnmasp	N-(2,2-diphenylethyl)	
			glycine	Nbhm
	D-N-methylcysteine	Dnmcys	N-(3,3-diphenylpropyl)	
		-	glycine	Nbhe

-9-

	D-N-methylglutamine	Dnmgln	N-(3-guanidinopropyl)	
			glycine	Narg
	D-N-methylglutamate	Dnmglu	N-(1-hydroxyethyl)glycine	Nthr
	D-N-methylhistidine	Dnmhis	N-(hydroxyethyl))glycine	Nser
5	D-N-methylisoleucine	Dnmile	N-(imidazolylethyl))	
			glycine	Nhis
	D-N-methylleucine	Dnmleu	N-(3-indolylyethyl)	
	•		glycine	Nhtrp
	D-N-methyllysine	Dnmlys	N-methyl-γ-aminobutyrate	Nmgabu
10	N-methylcyclohexylalanine	Nmchexa	D-N-methylmethionine	Dnmmet
	D-N-methylornithine	Dnmorn	N-methylcyclopentylalanine	Nmcpen
	N-methylglycine	Nala	D-N-methylphenylalanine	Dnmphe
	N-methylaminoisobutyrate	Nmaib	D-N-methylproline	Dnmpro
	N-(1-methylpropyl)glycine	Nile	D-N-methylserine	Dnmser
15	N-(2-methylpropyl)glycine	Nleu	D-N-methylthreonine	Dnmthr
	D-N-methyltryptophan	Dnmtrp	N-(1-methylethyl)glycine	Nval
	D-N-methyltyrosine	Dnmtyr	N-methyla-napthylalanine	Nmanap
	D-N-methylvaline	Dnmval	N-methylpenicillamine	Nmpen
	γ-aminobutyric acid	Gabu	N-(p-hydroxyphenyl)glycine	Nhtyr
20	L-t-butylglycine	Tbug	N-(thiomethyl)glycine	Ncys
	L-ethylglycine	Etg	penicillamine	Pen
	L-homophenylalanine	Hphe	L-a-methylalanine	Mala
	L-α-methylarginine	Marg	L-α-methylasparagine	Masn
	L-α-methylaspartate	Masp	L-α-methyl-t-butylglycine	Mtbug
25	L-a-methylcysteine	Mcys	L-methylethylglycine	Metg
	L-α-methylglutamine	Mgln	L-α-methylglutamate	Mglu
	L-a-methylhistidine	Mhis	L-α-methylhomo	
			phenylalanine	Mhphe
	L-a-methylisoleucine	Mile	N-(2-methylthioethyl)	
30			glycine	Nmet
	L-a-methylleucine	Mleu	L-α-methyllysine	Mlys

	L-α-methylmethionine	Mmet	L-α-methylnorleucine	Mnle
	L-\(\alpha\)-methylnorvaline	Mnva	L-α-methylornithine	Morn
	L-α-methylphenylalanine	Mphe	L-α-methylproline	Mpro
	L-α-methylserine	Mser	L-a-methylthreonine	Mthr
5	L-α-methyltryptophan	Mtrp	L-α-methyltyrosine	Mtyr
	L-α-methylvaline	Mval	L-N-methylhomo	
			phenylalanine	Nmhphe
	N-(N-(2,2-diphenylethyl)		N-(N-(3,3-diphenylpropyl)	
	carbamylmethyl)glycine	Nnbhm;	carbamylmethyl)glycine	Nnbhe
10	1-carboxy-1-(2,2-diphenyl-			
	ethylamino)cyclopropane	Nmbc		

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides which encodes, or are complementary to a sequence which encodes, a mammalian α -N-acetylglucosaminidase or fragment or derivative thereof or its like molecule.

Preferably, the mammal is a human, livestock animal, companion animal, wild animal or laboratory test animal (e.g. rabbit, rat, mouse or guinea pig). Most preferably, the mammal is a human. Conveniently, the α -N-acetylglucosaminidase is isolatable from the liver, kidney or placenta. However, the present invention extends to all mammalian α -N-acetylglucosaminidase enzymes and from any anatomical or cellular source and/or any biological fluid source, such as but not limited to plasma, serum, cell extract or lymph fluid.

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Although a preferred embodiment of the present invention contemplates the use of human α -N-acetylglucosaminidase or genomic or recombinant (e.g. cDNA) genetic sequences encoding same in the investigation, diagnosis and/or treatment of human subjects (i.e. homologous system), one skilled in the art will appreciate that the enzyme or genetic sequences encoding same from a non-human animal may also be useful. Such a heterologous system is encompassed by the present invention.

The term "nucleic acid molecule" as used herein shall be taken to refer to any RNA or DNA (eg. cDNA) molecule, whether single-stranded or double-stranded or in a linear or covalently-closed form. The nucleic acid molecule may also be DNA corresponding to the entire genomic gene or a substantial portion thereof or a fragment or derivative thereof.

The nucleic acid molecule of the present invention may constitute solely the nucleotide sequence encoding α -N-acetylglucosaminidase or a α -N-acetylglucosaminidase-like molecule or may be part of a larger nucleic acid molecule. Accordingly, the present invention extends to the isolated genomic α -N-acetylglucosaminidase gene. The non-

translated sequences in a larger nucleic acid molecule may include vector, transcriptional and/or translational regulatory sequences, promoter, terminator, enhancer, replication or signal sequences or non-coding regions (eg intron sequences) of an isolated genomic gene.

- 5 Reference herein to a "gene" is to be taken in its broadest context and includes:
 - (i) a classical genomic gene consisting of transcriptional and/or translational regulatory sequences and/or a coding region and/or non-translated sequences (i.e. introns, 5'- and 3'- untranslated sequences);
 - (ii) mRNA or cDNA corresponding to the coding regions (i.e. exons) optionally comprising 5'- or 3'-untranslated sequences of the gene; or
 - (iii) synthetic, amplified DNA fragments or other recombinant nucleic acid molecules produced *in vitro* and comprising all or a part of the coding region and/or 5'- or 3'-untranslated sequences of the gene.
- The term "gene" is also used to describe synthetic or fusion molecules encoding all or part of a functional product. A functional product is one which comprises a sequence of nucleotides or is complementary to a sequence of nucleotides which encodes a functional polypeptide, in particular a polypeptide having the catalytic activity of α-N-acetylglucosaminidase or a homologue, analogue or derivative thereof.

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For the present purpose, "homologues" of a nucleotide sequence shall be taken to refer to an isolated nucleic acid molecule which is substantially the same as the nucleic acid molecule of the present invention or its complementary nucleotide sequence, notwithstanding the occurrence within said sequence, of one or more nucleotide substitutions, insertions, deletions, or rearrangements.

"Analogues" of a nucleotide sequence set forth herein shall be taken to refer to an isolated nucleic acid molecule which is substantially the same as a nucleic acid molecule of the present invention or its complementary nucleotide sequence, notwithstanding the occurrence of any non-nucleotide constituents not normally present in said isolated nucleic acid molecule, for example carbohydrates, radiochemicals including

radionucleotides, reporter molecules such as, but not limited to DIG, alkaline phosphatase or horseradish peroxidase, amongst others.

"Derivatives" of a nucleotide sequence set forth herein shall be taken to refer to any isolated nucleic acid molecule which contains significant sequence similarity to said sequence or a part thereof. Generally, the nucleotide sequence of the present invention may be subjected to mutagenesis to produce single or multiple nucleotide substitutions, deletions and/or insertions. Nucleotide insertional derivatives of the nucleotide sequence of the present invention include 5' and 3' terminal fusions as well as intra-sequence insertions of single or multiple nucleotides or nucleotide analogues. Insertional nucleotide sequence variants are those in which one or more nucleotides or nucleotide analogues are introduced into a predetermined site in the nucleotide sequence of said sequence, although random insertion is also possible with suitable screening of the resulting product being performed. Deletional variants are characterised by the removal of one or more nucleotides from the nucleotide sequence. Substitutional nucleotide variants are those in which at least one nucleotide in the sequence has been removed and a different nucleotide or nucleotide analogue inserted in its place.

Preferably, a homologue, analogue or derivative of an α -N-acetylglucosaminidase gene according to any embodiments described herein, comprises a sequence of nucleotides of at least 10 contiguous nucleotides derived from SEQ ID NO:1 or SEQ ID NO:3 or a complementary strand thereof, wherein the sequence of said homologue, analogue or derivative is at least 40% identical to SEQ ID NO:1 or SEQ ID NO:3 or a complementary strand thereof or wherein said homologue, analogue or derivative is capable of hybridising to said sequence under at least low stringency hybridisation conditions.

For the purposes of nomenclature, the nucleotide sequence set for in SEQ ID NO: 1 relates to the cDNA encoding the human α -N-acetylglucosaminidase enzyme.

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The nucleotide sequence set forth in SEQ ID NO:3 relates to the genomic gene equivalent

of the cDNA encoding the human liver α -N-acetylglucosaminidase enzyme. Those skilled in the art will be aware that the specific exon sequences described in SEQ ID NO:3 correspond to the coding regions of the α -N-acetylglucosaminidase gene, said exon regions further comprising the entire open reading frame of the cDNA sequence set forth in SEQ ID NO:1, when aligned in a head-to-tail configuration. The intron sequences of SEQ ID NO:3, which correspond to non-coding regions of the gene which are spliced from the primary transcription product thereof, although not explicitly defined, may be readily deduced by those skilled in the art, when provided with the exon sequence data provided in the nucleotide sequence listing.

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The nucleotide sequence of the present invention may correspond to the sequence of the naturally-occurring α -N-acetylglucosaminidase gene or may comprise a homologue, analogue or derivative thereof which contains single or multiple nucleotide substitutions, deletions and/or additions. All such homologues, analogue or derivatives encode α -N-acetylglucosaminidase or α -N-acetylglucosaminidase-like molecules or a homologue, analogue or derivative thereof as contemplated by the present invention. The length of the nucleotide sequence may vary from a few bases, such as in nucleic acid probes or primers, to a full length sequence.

The present invention is particularly directed to the nucleic acid in cDNA form and particularly when inserted into an expression vector. The expression vector may be replicable in a eukaryotic or prokaryotic cell and may either produce mRNA or the mRNA may be subsequently translated into α-N-acetylglucosaminidase or like molecule. Particularly preferred eukaryotic cells include CHO cells but may be in any other suitable mammalian cells or cell lines or non-mammalian cells such as yeast or insect cells.

In an alternative embodiment, the present invention provides a nucleic acid molecule comprising a sequence of nucleotides which encodes or are complementary to a sequence which encodes a polypeptide capable of hydrolysing the α -N-acetylglucosamine residues from the non-reducing terminus of heparan sulphate and heparin fragments and wherein said nucleotide sequence is capable of hybridising under at least low stringency conditions

to a nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID NO:3 or a homologue, analogue or derivative thereof.

A second aspect of the invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides which is capable of hybridising under at least low stringency conditions to a nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID NO:3 or a complementary strand or a homologue, analogue or derivative thereof.

Preferably, hybridisation is possible under at least medium stringent conditions. More preferably, hybridisation is possible under high stringent conditions.

For the purposes of defining the level of stringency, reference can conveniently be made to Sambrook *et al* (1989) or Ausubel *et al* (1987) which are herein incorporated by reference.

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A low stringency is defined herein as being a hybridisation and/or wash carried out in 4-6X SSC/0.1-0.5% w/v SDS at 37-45°C for 2-3 hours. A medium stringency hybridisation and/or wash is carried out in 1-4X SSC/0.25-0.5% w/v SDS at ≥ 45°C for 2-3 hours and a high stringency hybridisation and/or wash is carried out 0.1-1X SSC/0.1% w/v SDS at 60°C for 1-3 hours.

Alternative conditions of stringency may be employed to those specifically recited herein. Generally, the stringency is increased by reducing the concentration of SSC buffer, and/or increasing the concentration of SDS and/or increasing the temperature of the hybridisation and/or wash. Those skilled in the art will be aware that the conditions for hybridisation and/or wash may vary depending upon the nature of the hybridisation membrane or the type of hybridisation probe used. Conditions for hybridisations and washes are well understood by one normally skilled in the art. For the purposes of clarification of parameters affecting hybridisation between nucleic acid molecules, reference is found in pages 2.10.8 to 2.10.16. of Ausubel et al. (1987), which is herein incorporated by reference.

Those skilled in the art will be aware that the nucleotide sequences set forth in SEQ ID NO:1 and SEQ ID NO:3 may be used to isolate the corresponding genes from other human tissues or alternatively, from the tissues or cells of other species, without undue experimentation. Means for the isolated of such related sequences will also be known to those skilled in the art, for example nucleic acid hybridisation, polymerase chain reaction, antibody screening of expression libraries, functional screening of expression libraries, or complementation of mutants, amongst others. The present invention is not to be limited by the source from which the specific gene sequences described herein have been isolated or by the means used to isolate said sequences.

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In one embodiment, a related genetic sequence comprising genomic DNA, or mRNA, or cDNA is contacted with a hybridisation effective amount of a genetic sequence which encodes α -N-acetylglucosaminidase, or its complementary nucleotide sequence or a homologue, analogue, derivative or functional part thereof, and then said hybridisation is detected using a suitable detection means.

The related genetic sequence may be in a recombinant form, in a virus particle, bacteriophage particle, yeast cell, animal cell, or a plant cell. Preferably, the related genetic sequence originates from an animal species or a human. More preferably, the related genetic sequence originates from a human.

Preferably, the genetic sequence which encodes α-N-acetylglucosaminidase (i.e probe or latter genetic sequence) comprises a sequence of nucleotides of at least 10 nucleotides, more preferably at least 20 nucleotides, even more preferably at least 50 nucleotides and even still more preferably at least 100 nucleotides derived from the nucleotide sequence set forth in SEQ ID NO: 1 or SEQ ID NO:3 or a complementary sequence or a homologue, analogue or derivative thereof.

Preferably, the detection means is a reporter molecule capable of giving an identifiable signal (e.g. a radioisotope such as ³²P or ³⁵S or a biotinylated molecule) covalently attached to the α-N-acetylglucosaminidase probe.

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In an alternative embodiment, the detection means is a polymerase chain reaction. According to this embodiment, two opposing non-complementary nucleic acid "primer molecules" of at least 10 nucleotides in length, more preferably at least 20 nucleotides in length, derived from the nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID NO:3 may be contacted with a nucleic acid "template molecule" and specific nucleic acid molecule copies of the template molecule amplified in a polymerase chain reaction.

The opposing primer molecules are selected such that they are each capable of hybridising to complementary strands of the same template molecule, wherein DNA polymerase-dependant DNA synthesis occurring from a first opposing primer molecule will be in a direction toward the second opposing primer molecule.

Accordingly, both primers hybridise to said template molecule such that, in the presence of a DNA polymerase enzyme, a cofactor and appropriate substrate, DNA synthesis occurs in the 5' to 3' direction from each primer molecule towards the position on the DNA where the other primer molecule is hybridised, thereby amplifying the intervening DNA.

Those skilled in the art are aware of the technical requirements of the polymerase chain reaction and are capable of any modifications which may be made to the reaction conditions. For example, of the polymerase chain reaction may be used in any suitable format, such as amplified fragment length polymorphism (AFLP), single-strand chain polymorphism (SSCP), amplification and mismatch detection (AMD), interspersed repetitive sequence polymerase chain reaction (IRS-PCR), inverse polymerase chain reaction (iPCR) and reverse transcription polymerase chain reaction (RT-PCR), amongst others, to isolate a related α -N-acetylglucosaminidase gene sequence or identify a mutation in an α -N-acetylglucosaminidase genetic sequence. Such variations of the polymerase chain reaction are discussed in detail by McPherson *et al* (1991), which is incorporated herein by reference. The present invention encompasses all such variations, the only requirement being that the final product of the reaction is an isolated nucleic acid molecule which is capable of encoding α -N-acetylglucosaminidase or a homologue,

analogue or derivative thereof.

In a preferred embodiment, the first primer molecule is preferably derived from the sense strand of a gene which encodes α -N-acetylglucosaminidase, in particular from the nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID NO:3 or a homologue, derivative or analogue thereof and the second primer molecule is preferably derived from the antisense strand of said gene.

Those skilled in the art will be aware that it is not essential to the performance of the invention that the primer molecules be derived from the same gene.

According to this embodiment, the nucleic acid primer molecule may further consist of a combination of any of the nucleotides adenine, cytidine, guanine, thymidine, or inosine, or functional analogues or derivatives thereof, capable of being incorporated into a polynucleotide molecule provided that it is capable of hybridising under at least low stringency conditions to the nucleic acid molecule set forth in SEQ ID NO:1 or SEQ ID NO:3 or a homologue, analogue or derivative thereof.

The nucleic acid primer molecules may further be each contained in an aqueous pool comprising other nucleic acid primer molecules. More preferably, the nucleic acid primer molecule is in a substantially pure form.

The nucleic acid template molecule may be in a recombinant form, in a virus particle, bacteriophage particle, yeast cell, animal cell, or a plant cell. Preferably, the related genetic sequence originates from a cell, tissue, or organ derived from an animal species or a human. More preferably, the related genetic sequence originates from a cell, tissue, or organ derived from a human.

Accordingly, a third aspect of the present invention extends to an isolated nucleic acid molecule which is at least 40% identical to the nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID NO:3 or to a complementary strand thereof or a homologue, analogue

or derivative thereof.

Preferably, the percentage identity to SEQ ID NO:1 or SEQ ID NO:3 is at least about 55%, still more preferably at least about 65%, yet still more preferably at least about 75-80% and even still more preferably at least about 85-95%.

In an even more preferred embodiment, the present invention provides an isolated nucleic acid molecule which is at least 40% identical to the nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID NO:3 or to a complementary strand thereof or a homologue, analogue or derivative thereof and is capable of hybridising under at least low stringency conditions to a nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID NO:3.

In a particularly preferred embodiment, the isolated nucleic acid molecule described herein is further capable of encoding a sequence of amino acids which is capable of carrying out the enzyme reaction catalysed by a α -N-acetylglucosaminidase enzyme.

The isolated nucleic acid molecule of the present invention is also useful for developing a genetic construct comprising a sense molecule, for the expression or over-expression of α -N-acetylglucosaminidase in prokaryotic or eukaryotic cells. Particularly preferred eukaryotic cells include CHO cells but may be in any other suitable mammalian cells or cell lines or non-mammalian cells such as yeast or insect cells.

The term "sense molecule" as used herein shall be taken to refer to an isolated nucleic acid molecule of the invention as described herein, which is provided in a format suitable for its expression to produce a recombinant polypeptide, when said sense molecule is introduced into a host cell.

In a particularly preferred embodiment, a sense molecule which encodes the α-N-acetylglucosaminidase comprises a sequence of nucleotides set forth in SEQ ID NO:1 or SEO ID NO:3 or a complementary strand, homologue, analogue or derivative thereof.

In a most particularly preferred embodiment, the sense molecule of the invention comprises the sequence of nucleotides set forth in SEQ ID NO:1 or a complementary strand, homologue, analogue or derivative thereof.

Those skilled in the art will be aware that expression of a sense molecule may require the nucleic acid molecule of the invention to be placed in operable connection with a promoter sequence to produce a "sense construct". The choice of promoter for the present purpose may vary depending upon the level of expression of the sense molecule required and/or the tissue-specificity or developmental-specificity of expression of the sense molecule which is required. The sense construct may further comprise a terminator sequence and be introduced into a suitable host cell where it is capable of being expressed to produce a recombinant polypeptide gene product.

In the context of the present invention, a sense molecule which corresponds to a genetic sequence or isolated nucleic acid molecule which encodes α -N-acetylglucosaminidase polypeptide or a homologue, analogue or derivative thereof, placed operably under the control of a suitable promoter sequence, is introduced into a cell using any suitable method for the transformation of said cell and said genetic sequence or isolated nucleic acid molecule is expressed therein to produce said polypeptide.

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The present invention clearly extends to genetic constructs designed to facilitate expression of any nucleic acid molecule described herein.

A genetic construct of the present invention comprises the foregoing sense molecule, placed operably under the control of a promoter sequence capable of regulating the expression of the said nucleic acid molecule in a prokaryotic or eukaryotic cell, preferably a mammalian cell such as a CHO cell, a yeast cell, insect cell or bacterial cell. The said genetic construct optionally comprises, in addition to a promoter and sense molecule, a terminator sequence.

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The term "terminator" refers to a DNA sequence at the end of a transcriptional unit

which signals termination of transcription. Terminators are 3'-non-translated DNA sequences containing a polyadenylation signal, which facilitates the addition of polyadenylate sequences to the 3'-end of a primary transcript. Terminators active in plant cells are known and described in the literature. They may be isolated from bacteria, fungi, viruses, animals and/or plants.

Reference herein to a "promoter" is to be taken in its broadest context and includes the transcriptional regulatory sequences of a classical genomic gene, including the TATA box which is required for accurate transcription initiation, with or without a CCAAT box sequence and additional regulatory elements (i.e. upstream activating sequences, enhancers and silencers) which alter gene expression in response to developmental and/or external stimuli, or in a tissue-specific manner. A promoter is usually, but not necessarily, positioned upstream or 5', of a structural gene, the expression of which it regulates. Furthermore, the regulatory elements comprising a promoter are usually positioned within 2 kb of the start site of transcription of the gene.

In the present context, the term "promoter" is also used to describe a synthetic or fusion molecule, or derivative which confers, activates or enhances expression of said sense molecule in a cell.

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Preferred promoters may contain additional copies of one or more specific regulatory elements, to further enhance expression of the sense molecule and/or to alter the spatial expression and/or temporal expression of said sense molecule. For example, regulatory elements which confer copper inducibility may be placed adjacent to a heterologous promoter sequence driving expression of a sense molecule, thereby conferring copper inducibility on the expression of said molecule.

Placing a sense molecule under the regulatory control of a promoter sequence means positioning the said molecule such that expression is controlled by the promoter sequence. Promoters are generally positioned 5' (upstream) to the genes that they control. In the construction of heterologous promoter/structural gene combinations it is

generally preferred to position the promoter at a distance from the gene transcription start site that is approximately the same as the distance between that promoter and the gene it controls in its natural setting, i.e., the gene from which the promoter is derived. As is known in the art, some variation in this distance can be accommodated without loss of promoter function. Similarly, the preferred positioning of a regulatory sequence element with respect to a heterologous gene to be placed under its control is defined by the positioning of the element in its natural setting, i.e., the genes from which it is derived. Again, as is known in the art, some variation in this distance can also occur.

- Examples of promoters suitable for use in genetic constructs of the present invention include viral, fungal, bacterial, animal and plant derived promoters capable of functioning in animal, human, yeast, insect or bacterial cells. The promoter may regulate the expression of the said molecule constitutively, or differentially with respect to the tissue in which expression occurs or, with respect to the developmental stage at which expression occurs, or in response to external stimuli such as physiological stresses, or plant pathogens, or metal ions, amongst others. Preferably, the promoter is capable of regulating expression of a sense molecule in a cell derived from an animal species or human.
- In a particularly preferred embodiment, the promoter is derived from the genomic gene encoding α-N-acetylglucosaminidase, preferably the human α-N-acetylglucosaminidase gene. In a more preferred embodiment, however, the promoter is derived from the nucleotide sequence set forth in SEQ ID NO:3 or is at least capable of hybridising to nucleotide residues 1 to 989 of SEQ ID NO:3 or at least 20 contiguous nucleotides derived therefrom.

In an even more particularly preferred embodiment, the promoter is the CMV promoter sequence or a promoter sequence derived therefrom.

An alternative embodiment of the invention is directed to a genetic construct comprising a promoter or functional derivative, part fragment, homologue, or analogue thereof,

derived from the α-N-acetylglucosaminidase genomic gene defined by SEQ ID NO: 3.

Preferably, said genetic construct further comprises the α-N-acetylglucosaminidase sequence defined by SEQ ID NO:1 placed in operably connection with said promoter.

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A further aspect of the present invention is directed to synthetic α -N-acetylglucosaminidase or like molecule.

The term "synthetic" as used herein shall be taken to include both recombinant and

chemically-synthesised molecules produced by the sequential addition of amino acid

residues or groups of amino acid residues in defined order.

In one embodiment, the invention relates to recombinant α-N-acetylglucosaminidase or

like molecule encoded by or expressed from the nucleic acid molecules as hereinbefore

15 described.

In another embodiment, the synthetic α -N-acetylglucosaminidase or like molecule

comprises a sequence of amino acids which is at least 40% identical to the amino acid

sequence set forth in any one of SEQ ID Nos:2, 4, 5 or 6.

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More preferably, the percentage identity is at least 60% and still more preferably at least

80% or 85-90%.

A particularly preferred embodiment of the present invention provides a synthetic α -N-

acetylglucosaminidase as hereinbefore defined which comprises a sequence of amino acids

substantially as set forth in any one of SEQ ID Nos:2, 4, 5 or 6 or a homologue, analogue

or derivative thereof.

For the purposes of nomenclature, the amino acid sequence set forth in SEQ ID NO:2

comprises the full-length translation product of the human α-N-acetylglucosaminidase gene

(i.e. hereinafter referred to as the "α-N-acetylglucosaminidase polypeptide" or "SEO ID

WO 97/19177 PCT/AU96/00747

- 24 -

NO:2") produced by expression of either the cDNA sequence defined by SEQ ID NO:1 or the genomic gene defined by SEQ ID NO:3. The α -N-acetylglucosaminidase polypeptide comprises at least seven potentially-glycosylated Asn residues, at positions 261, 272, 435, 503, 513, 526 and 532. Furthermore, the amino acid sequence of the α -N-acetylglucosaminidase polypeptide may comprise a signal peptide of approximately 23 amino acid residues in length, with a probable site for signal peptide peptidase cleavage occurring between Gly₂₃ and Asp₂₄.

The amino acid sequences set forth in SEQ ID Nos:4-6 relate to N-terminal and internal (i.e. CNBr) amino acid sequences derived from human α-N-acetylglucosaminidase, purified as described in Example 1. As described in Example 2, the purified form of the enzyme comprises two polypeptides having approximate molecular weights of 82 and 77 kDa. The sequence set forth in SEQ ID NO:4 relates to the N-terminal sequence of the 82 kDa polypeptide, while SEQ ID NO:5 relates to the N-terminal sequence of the 77 kDa polypeptide. Furthermore, SEQ ID NO:4 comprises amino acids residues 24-43 of SEQ ID NO:2, while SEQ ID NO:5 comprises amino acid residues 59-76 of SEQ ID NO:2.

The amino acid sequence defined by SEQ ID NO:6 relates to the CNBr-cleaved peptide of purified human α -N-acetylglucosaminidase. This amino acid sequence aligns with amino acid residues 540-554 of the α -N-acetylglucosaminidase polypeptide (SEQ ID NO:2).

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In the present context, "homologues" of a polypeptide refer to those polypeptides, enzymes or proteins which have a similar α -N-acetylglucosaminidase enzyme activity, notwithstanding any amino acid substitutions, additions or deletions thereto. A homologue may be isolated or derived from the same or another animal species.

Furthermore, the amino acids of a homologous polypeptide may be replaced by other amino acids having similar properties, for example hydrophobicity, hydrophilicity, hydrophobic moment, charge or antigenicity, and so on.

"Analogues" encompass α -N-acetylglucosaminidase polypeptides and peptide derivatives thereof notwithstanding the occurrence of any non-naturally occurring amino acid analogues therein.

The term "derivative" in relation to the polypeptides of the invention refer to mutants, parts or fragments of a functional molecule. Derivatives include modified peptides in which ligands are attached to one or more of the amino acid residues contained therein, such as carbohydrates, enzymes, proteins, polypeptides or reporter molecules such as radionuclides or fluorescent compounds. Glycosylated, fluorescent, acylated or alkylated forms of the subject peptides are particularly contemplated by the present invention. Additionally, derivatives of a polypeptide may comprise fragments or parts of an amino acid sequence disclosed herein and are within the scope of the invention, as are homopolymers or heteropolymers comprising two or more copies of the subject polypeptides. Procedures for derivatizing peptides are well-known in the art.

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Accordingly, this aspect of the present invention is directed to any proteinaceous molecule comprising an amino acid sequence corresponding to the full length mammalian α -N-acetylglucosaminidase enzyme or to a like molecule. The like molecule, therefore, comprises parts, derivatives and/or portions of the α -N-acetylglucosaminidase enzyme whether functional or not.

Preferably, the mammal is human but may be of non-human origin as contemplated above.

The synthetic or recombinant α -N-acetylglucosaminidase of the present invention may comprise an amino acid sequence corresponding to the naturally occurring amino acid sequence or may contain single or multiple amino acid substitutions, deletions and/or additions. The length of the amino acid sequence may range from a few residues to a full length molecule.

Amino acid substitutions are typically of single residues. Amino acid insertions will usually be in the order of about 1-10 amino acid residues and deletions will range from

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about 1-20 residues. Preferably, deletions or insertions are made in adjacent pairs, i.e. a deletion of two residues or insertion of two residues.

Amino acid insertional derivatives of α -N-acetylglucosaminidase of the present invention include amino and/or carboxyl terminal fusions as well as intra-sequence insertions of single or multiple amino acids. Insertional amino acid sequence variants are those in which one or more amino acid residues are introduced into a predetermined site in the protein although random insertion is also possible with suitable screening of the resulting product. Deletional variants are characterised by the removal of one or more amino acids from the sequence. Substitutional amino acid variants are those in which at least one residue in the sequence has been removed and a different residue inserted in its place. Typical substitutions are those made in accordance with the following Table 2:

The amino acid variants referred to above may readily be made using peptide synthetic techniques well known in the art, such as solid phase peptide synthesis (Merrifield synthesis) and the like, or by recombinant DNA manipulations. Techniques for making substitution mutations at predetermined sites in DNA having known or partially known sequence are well known and include, for example, M13 mutagenesis. The manipulation of DNA sequence to produce variant proteins which manifest as substitutional, insertional or deletional variants are conveniently elsewhere described such as Sambrook et al, 1989 Molecular Cloning: A Laboratory Manual Cold Spring Harbor Laboratories, Cold Spring Harbor, NY.

The derivatives or like molecules include single or multiple substitutions, deletions and/or additions of any component(s) naturally or artificially associated with the α -N-acetylglucosaminidase enzyme such as carbohydrate, lipid and/or other proteinaceous moieties. For example, the present invention extends to glycosylated and non-glycosylated forms of the molecule. All such molecules are encompassed by the expression "mutants", "derivatives", "fragments", "portions" and "like" molecules. These molecules may be active or non-active and may contain specific regions, such as a catalytic region. Particularly, preferred derivative molecules include those with altered glycosylation

patterns relative to the naturally occurring molecule. Even more particularly, the recombinant molecule is more highly glycosylated than the naturally occurring molecule. Such highly glycosylated derivatives may have improved take-up properties and enhanced half-lives.

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As indicated in the Examples, the molecular weight of purified human α -N-acetylglucosaminidase (i.e. 82kDa and 77kDa) and recombinant mammalian α -N-acetylglucosaminidase produced in CHO cells (i.e. 89 kDa and 79 kDa) are greater than the deduced molecular weight of the α -N-acetylglucosaminidase polypeptide set forth in SEQ ID No:2 (i.e. 70 kDa), suggesting that the purified and recombinant polypeptide are post-translationally modified. The data presented in Example 8 indicate further that the recombinant α -N-acetylglucosaminidase enzyme produced in CHO cells, at least, is glycosylated and that the difference in molecular weight determined for the recombinant polypeptides and the polypeptide of SEQ ID No: 2 is due almost entirely to glycosylated recombinant polypeptide by CHO cells. As shown in Example 9, the glycosylated recombinant α -N-acetylglucosaminidase polypeptide exhibits enzymatic activity.

The present invention also extends to synthetic α -N-acetylglucosaminidase or like molecules when fused to other proteinaceous molecules. The latter may include another enzyme, reporter molecule, purification site or an amino acid sequence which facilitates transport of the molecule out of a cell, such as a signal sequence.

The present invention extends further to post-translational modifications to the α -N-acetylglucosaminidase enzyme. The modifications may be made to the naturally occurring enzyme or following synthesis by recombinant techniques. The modifications may be at the structural level or at, for example, the electrochemical level such as modifying net charge or structural conformation of the enzyme.

Such modification may be important to facilitate entry or penetration of the enzyme into selected tissues such as cartilage or blood brain barriers or to increase circulation half-life.

WO 97/19177

Analogues of α -N-acetylglucosaminidase contemplated herein include, but are not limited to, modifications to side chains, incorporation of unnatural amino acids and/or their derivatives during peptide synthesis and the use of crosslinkers and other methods which impose conformational constraints on the enzyme.

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Examples of side chain modifications contemplated by the present invention include modifications of amino groups such as by reductive alkylation by reaction with an aldehyde followed by reduction with NaBH₄; amidination with methylacetimidate; acylation with acetic anhydride; carbamoylation of amino groups with cyanate; trinitrobenzylation of amino groups with 2, 4, 6-trinitrobenzene sulphonic acid (TNBS); acylation of amino groups with succinic anhydride and tetrahydrophthalic anhydride; and pyridoxylation of lysine with pyridoxal-5'-phosphate followed by reduction with NaBH₄.

The guanidino group of arginine residues may be modified by the formation of heterocyclic condensation products with reagents such as 2,3-butanedione, phenylglyoxal and glyoxal.

The carboxyl group may be modified by carbodiimide activation via O-acylisourea formation followed by subsequent derivatisation, for example, to a corresponding amide.

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Sulphydryl groups may be modified by methods such as carboxymethylation with iodoacetic acid or iodoacetamide; performic acid oxidation to cysteic acid; formation of a mixed disulphides with other thiol compounds; reaction with maleimide, maleic anhydride or other substituted maleimide; formation of mercurial derivatives using 4-chloromercuribenzoate, 4-chloromercuriphenylsulphonic acid, phenylmercury chloride, 2-chloromercuric-4-nitrophenol and other mercurials; carbamoylation with cyanate at alkaline pH.

Tryptophan residues may be modified by, for example, oxidation with Nbromosuccinimide or alkylation of the indole ring with 2-hydroxy-5-nitrobenzyl bromide
or sulphenyl halides. Tyrosine residues on the other hand, may be altered by nitration with

tetranitromethane to form a 3-nitrotyrosine derivative.

Modification of the imidazole ring of a histidine residue may be accomplished by alkylation with iodoacetic acid derivatives or N-carbethoxylation with diethylpyrocarbonate.

Examples of incorporating unnatural amino acids and derivatives during peptide synthesis include, but are not limited to, use of norleucine, 4-amino butyric acid, 4-amino-3-hydroxy-5-phenylpentanoic acid, 6-aminohexanoic acid, t-butylglycine, norvaline, phenylglycine, ornithine, sarcosine, 4-amino-3-hydroxy-6-methylheptanoic acid, 2-thienyl alanine and/or D-isomers of amino acids. Non-naturally occurring amino acids contemplated by the present invention are incorporated herein, as Table 3.

Crosslinkers can be used, for example, to stabilise 3D conformations, using homobifunctional crosslinkers such as the bifunctional imido esters having $(CH_2)_n$ spacer groups with n=1 to n=6, glutaraldehyde, N-hydroxysuccinimide esters and hetero-bifunctional reagents which usually contain an amino-reactive moiety such as N-hydroxysuccinimide and another group specific-reactive moiety such as maleimido or dithio moiety (SH) or carbodiimide (COOH). In addition, the enzyme could be conformationally constrained by, for example, incorporation of C_a and N_a -methylamino acids, introduction of double bonds between C_a and C_b atoms of amino acids and the formation of cyclic peptides or analogues by introducing covalent bonds such as forming an amide bond between the N and C termini, between two side chains or between a side chain and the N or C terminus.

25 Electrochemical modifications of α-N-acetylglucosaminidase include interaction with polylysine or polyethylene glycol or other agent which effects an overall change to the net charge of the enzyme.

Advantageously, the recombinant α -N-acetylglucosaminidase is a biologically pure preparation meaning that it has undergone some purification away for other proteins and/or non-proteinaceous material. The purity of the preparation may be represented as at least

40% of the enzyme, preferably at least 60%, more preferably at least 75%, even more preferably at least 85% and still more preferably at least 95% relative to non-α-N-acetylglucosaminidase material as determined by weight, activity, amino acid homology or similarity, antibody reactivity or other convenient means.

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Particularly preferred methods for the preparation and purification of recombinant α -N-acetylglucosaminidase are provided in Examples 7 and 8.

Those skilled in the art will be aware of the means of purifying a synthetic or recombinant α-N-acetylglucosaminidase from several sources without undue experimentation and for expressing the degree of purity of such a purified preparation of the enzyme.

The present invention further contemplates antibodies to α -N-acetylglucosaminidase and preferably synthetic α -N-acetylglucosaminidase or like molecule. The antibodies may be polyclonal or monoclonal, naturally occurring or synthetic (including recombinant, fragment or fusion forms). Such antibodies will be useful in developing immunoassays for α -N-acetylglucosaminidase and for identifying additional genetic sequences which are capable of expressing α -N-acetylglucosaminidase polypeptides or homologues, analogues or derivatives thereof.

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Both polyclonal and monoclonal antibodies are obtainable by immunisation with an appropriate synthetic or recombinant gene product, or epitope, or peptide fragment of a gene product, in particular a α -N-acetylglucosaminidase polypeptide or a homologue, analogue or derivative thereof.

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Alternatively, fragments of antibodies may be used, such as Fab fragments. The present invention extends further to encompass recombinant and synthetic antibodies and to antibody hybrids. A "synthetic antibody" is considered herein to include fragments and hybrids of antibodies.

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A further aspect of the present invention contemplates a method of screening for mutations

or other abberations in the α -N-acetylglucosaminidase gene in a human or animal patient. Such a method may be accomplished in a number of ways including isolating a source of DNA to be tested or mRNA therefrom and hybridising thereto a nucleic acid molecule as hereinbefore described. Generally, the nucleic acid is probe or primer size and polymerase chain reaction is a convenient means by which to analyse the RNA or DNA. Other suitable assays include the ligation chain reaction and the strand displacement amplification methods. The α -N-acetylglucosaminidase sequence can also be determined and compared to the naturally occurring sequence. Such methods may be useful in adults and children and may be adapted for a pre-natal test. The DNA to be tested includes a genomic sample carrying the α -N-acetylglucosaminidase gene, a cDNA clone and/or amplification product.

In accordance with this aspect of the present invention there is provided a method for screening for abberations in the α -N-acetylglucosaminidase gene including the absence of such a gene or a portion or a substantial portion thereof comprising isolating a sample of DNA or mRNA corresponding to a region of said DNA and contacting same with an oligonucleotide probe capable of hybridising to one or more complementary sequences within the α -N-acetylglucosaminidase gene and then detecting the hybridisation, the extent of hybridisation or the absence of hybridisation.

- Alternatively, the probe is a primer and capable of directing amplification of one or more regions of said α-N-acetylglucosaminidase gene and the amplification products and/or profile of amplification products is compared to an individual carrying the full gene or to a reference date base.
- 25 Conveniently, the amplification products are sequenced to determine the presence or absence of the full gene.

The present invention extends to the use of any and all DNA-based or nucleic acid-based hybridisation and/or polymerase chain reaction formats as described herein, for the diagnosis of a disorder involving the α-N-acetylglucosaminidase gene in a human or animal patient.

The present invention further extends to a method of treating patients suffering from α -N-acetylglucosaminidase deficiency, such as in MPS-IIIB, said method comprising administering to said patient an effective amount of α -N-acetylglucosaminidase or active like form thereof.

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Preferably, the α -N-acetylglucosaminidase is in recombinant form. Such a method is referred to as "enzyme therapy". Alternatively, gene therapy can be employed including introducing an active gene (i.e. a nucleic acid molecule as hereinbefore described) or to parts of the gene or other sequences which facilitate expression of a naturally occurring α -N-acetylglucosaminidase gene.

Administration of α -N-acetylglucosaminidase for enzyme therapy may be by oral, intravenous, suppository, intraperitoneal, intramuscular, intranasal, intradermal or subcutaneous administration or by infusion or implantation. The α -N-acetylglucosaminidase is preferably as hereinbefore described including active mutants or derivatives thereof and glycosylation variants thereof. Administration may also be by way of gene therapy including expression of the gene by inclusion of the gene in viral vectors which are introduced into the animal (e.g. human) host to be treated. Alternatively, the gene may be expressed in a bacterial host which is then introduced and becomes part of the bacterial flora in the animal to be tested.

Still yet another aspect of the present invention is directed to a pharmaceutical composition comprising synthetic (e.g. recombinant) α -N-acetylglucosaminidase or like molecule, including active derivatives and fragments thereof, alone or in combination with other active molecules. Such other molecules may act synergistically with the enzyme or facilitates its entry to a target cell. The composition will also contain one or more pharmaceutically acceptable carriers and/or diluents. The composition may alternatively comprise a genetic component useful in gene therapy.

30 The active ingredients of the pharmaceutical composition comprising the synthetic or recombinant α-N-acetylglucosaminidase or mutants or fragments or derivatives thereof are

contemplated to exhibit excellent activity in treating patients with a deficiency in the enzyme when administered in an amount which depends on the particular case. The variation depends, for example, on the patient and the α -N-acetylglucosaminidase used. For example, from about 0.5 ug to about 20 mg of enzyme per animal body or, depending on the animal and other factors, per kilogram of body weight may be administered. Dosage regima may be adjusted to provide the optimum therapeutic response. For example, several divided doses may be administered daily, weekly, monthly or in other suitable time intervals or the dose may be proportionally reduced as indicated by the exigencies of the situation. Accordingly, alternative dosages in the order of 1.0 μ g to 15 mg, 2.0 μ g to 10 mg or 10µg to 5mg may be administered in a single or as part of multiple doses. The active compound may be administered in a convenient manner such as by the oral, intravenous (where water soluble), intramuscular, subcutaneous, intranasal, intradermal or suppository routes or implanting (eg using slow release molecules). Depending on the route of administration, the active ingredients which comprise a synthetic (e.g. recombinant) a-N-acetylglucosaminidase or fragments, derivatives or mutants thereof may be required to be coated in a material to protect same from the action of enzymes, acids and other natural conditions which may inactivate said ingredients. For example, the low lipophilicity of a-N-acetylglucosaminidase will allow it to be destroyed in the gastrointestinal tract by enzymes capable of cleaving peptide bonds and in the stomach by acid hydrolysis. In order to administer the vaccine by other than parenteral administration, the enzyme will be coated by, or administered with, a material to prevent its inactivation. For example, the enzyme may be administered in an adjuvant, co-administered with enzyme inhibitors or in liposomes. Adjuvant is used in its broadest sense and includes any immune stimulating compound such as interferon. Adjuvants contemplated herein include resorcinols, nonionic surfactants such as polyoxyethylene oleyl ether and n-hexadecyl polyethylene ether. Conveniently, the adjuvant is Freund's Complete or Incomplete Adjuvant. Enzyme inhibitors include pancreatic trypsin inhibitor, diisopropylfluorophosphate (DEP) and trasylol. Liposomes include water-in-oil-in-water emulsions as well as conventional liposomes.

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The active compound may also be administered in dispersions prepared in glycerol, liquid

polyethylene glycols, and/or mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of superfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thirmerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions are prepared by incorporating the active compound in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterilized active ingredient(s) into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and the freeze-drying technique which yield a powder of the active ingredient plus any additional desired ingredient from previously sterile-filtered solution thereof.

When the \$\alpha\$-N-acetylglucosaminidase of the present invention is suitably protected as described above, the composition may be orally administered, for example, with an inert diluent or with an assimilable edible carrier, or it may be enclosed in hard or soft shell gelatin capsule, or it may be compressed into tablets, or it may be incorporated directly with the food of the diet. For oral therapeutic administration, the active compound may be incorporated with excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, suspensions, syrups, wafers, and the like. Such compositions and preparations should contain at least 1% by weight of active compound. The percentage of the compositions and preparations may, of course, be varied and may conveniently be between about 5 to about 80% of the weight of the unit. The amount of active compound in the vaccine compositions is such that a suitable dosage will be obtained. Preferred compositions or preparations according to the present invention are prepared, so that an oral dosage unit form contains between about 0.5 ug and 20 mg of active compound.

The tablets, troches, pills, capsules and the like may also contain the following: a binder such as gum gragacanth, acacia, corn starch or gelatin; excipients such as dicalcium phosphate; a disintegrating agent such as corn starch, potato starch, alginic acid and the like; a lubricant such as magnesium stearate; and a sweetening agent such a sucrose, lactose or saccharin may be added or a flavoring agent such as peppermint, oil of wintergreen, or cherry flavouring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar or both. A syrup or elixir may contain the active compound, sucrose as a sweetening agent, methyl and propylparabens as preservatives, a dye and flavoring such as cherry or orange flavor. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active compound may be incorporated into sustained-release reparations and formulations.

As used herein "pharmaceutically acceptable carriers and/or diluents" include any and all solvents, dispersion media, aqueous solutions, coatings, antibacterial and antifungal agents,

isotonic and absorption delaying agents, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, use thereof in the pharmaceutical compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

The present invention further relates to the use of α-N-acetylglucosaminidase or active fragment, mutant or derivative thereof in the manufacture of a medicament for the treatment of patients suffering from a deficiency in the naturally occurring enzyme (e.g. MPS-IIIB).

The present invention is further described with reference to the following non-limiting Examples.

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EXAMPLE 1

Purification of α-N-Acetylglucosaminidase

α-N-acetylglucosaminidase was purified according to the method described in Weber et al. (1996). Enzyme was purified to homogeneity from human placenta. Evidence of purity is shown following SDS/PAGE which is represented in Figure 1. All samples were reduced with dithiothreitol prior to electrophoresis.

EXAMPLE 2

Characterisation of α-N-Acetylglucosaminidase

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Results presented in Figure 1 show two polypeptides of about 82kDa and 77 kDa molecular weight, which correspond to α-N-acetylglucosaminidase polypeptides purified from human placenta according to Example 1.

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EXAMPLE 3

Amino Acid Sequence Determination

- 37 -

The N-terminal amino acid sequences the 77 kDA and 82 kDa α-N-acetylglucosaminidase polypeptides, in addition to the amino acid sequence of an internal CNBr cleavage product of these peptides, were determined using the methods of Weber et al. (1996).

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The amino acid sequences are shown in Table 4.

- 38 -

TABLE 4

N-Terminal amino acid sequences (SEQ ID NO:4 and SEQ ID No:5) and CNBr peptide sequence (SEQ ID No:6) determined from purified human α-N-Acetylglucosaminidase

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olypeptide 82 kDa	DEAREAAAVRALVARLLGPG
polypeptide 77 kDa	KPGLDTYSLGGGGAAX¹ VR
CNBr peptide	WRLLLTSAPSLX ¹ TX ¹ P

X¹ no residue could be identified for this position, indicating that this residue could be phosphorylated or glycosylated.

EXAMPLE 4

Cloning of a-N-Acetylglucosaminidase cDNA

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Oligonucleotide probes were prepared based on the partial amino acid sequences obtained for the purified α -N-acetylglucosaminidase polypeptides (Example 3). The probes were subsequently used to screen a human peripheral blood leukocyte cDNA library. An approximately 2.6 kbp cDNA clone was isolated encoding most of the sequence of human α -N-acetylglucosaminidase (SEQ ID NO:1).

The remaining α -N-acetylglucosaminidase coding sequence was obtained from the nucleotide sequence of the corresponding genomic gene (SEQ ID NO:3), isolated by hybridisation to a human chromosome 17 library (Weber *et.al.* 1996).

The complete open reading frame is 2232 nucleotides long and encodes a 743 (plus stop codon) amino acid protein. The predicted molecular mass of the longest mature protein (minus the 23 amino acid N-terminal signal peptide) is about 79,622 daltons.

The amino acid sequence of α-N-acetylglucosaminidase is shown in SEQ ID NO:2. The deduced molecular weight of the desired amino acid sequence of α-N-acetylglucosaminidase is approximately 70kDa. The probable site of signal peptide peptidase cleavage is between amino acids 23 and 24. There are seven potential N-glycosylation sites in the sequence.

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The nucleotide sequence of the corresponding α -N-acetylglucosaminidase genomic gene (SEQ ID No:3) comprises 10380 bp including 889 bp of 5' upstream sequence corresponding to at least at part of the α -N-acetylglucosaminidase promoter sequence, in addition to the nucleotide sequences of introns I, II, II, IV, V, in addition to 1326 bp of 3'-untranslated sequence.

EXAMPLE 5

Construction of an expression vector comprising the α-N-Acetylglucosaminidase cDNA sequence

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The cDNA insert of λ clone pbl 33, containing bases 107 to 2575 of the α -N-acetylglucosaminidase cDNA was excised with EcoRI and subcloned into pBluescript II SK-(Stratagene). A 178 bp XmaI fragment (bases 1 to 178 of the α -N-acetylglucosaminidase cDNA) from cosmid sub-clone 6.3, containing the start codon, was cloned into the pBluescript subclone to produce a full-length cDNA sequence in addition to 101 bp of 5' non-translated sequence as well as 245 bp of 3' non-translated region including the polyadenylation-site, the polyA-tail and linkerDNA. The full length cDNA was directionally cloned into the pCDNA3 expressionvector (Invitrogen) via the EcoRI and BamHI sites.

Expression of recombinant α-N-acetylglucosaminidase

Chinese Hamster Ovary (CHO) cells were transfected with expression vector using the DOTAP transfection reagent (Boehringer Mannheim) according to the manufacturers instructions. Cells were grown in Ham's F12 medium, 10% (v/v) fetal calf serum, penicillin and streptomycin sulfate at 100 μ g/ml each. Cells were grown in nonselective medium for 48 h and then incubated in medium containing 750 μ g/ml G418 sulfate (Geniticin) until resistant colonies emerged.

Single cell clones were grown up and 26 of them were tested for expression of recombinant α-N-acetylglucosaminidase with a fluorogenic α-N-acetylglucosaminidase substrate. (i.e. N-acetylglucosamine α-linked to 4-methylumbelliferone)

EXAMPLE 7

Large scale α-N-acetylglucosaminidase production

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2 g of Cytodex 2 microcarrier beads were swollen in 250 ml of PBS for 3 h at 37°C with three changes of PBS and then autoclaved for 15 min at 120°C (wet cycle). The beads were then rinsed with sterile growth medium (Coons/DMEM, 10% v/v fetal calf serum, penicillin and streptomycin sulfate at 100 μ g/ml each and 0.1% w/v Pluronic F68) and transferred into a Techne stirrer culture flask. The microcarrier beads were inoculated with seven confluent 175 flasks of the cell clone showing the highest expression of recombinant α -N-acetylglucosaminidase. Growth medium was added up to 200 ml and the culture incubated with a stirrer speed of 20 rpm to achieve an even distribution of cells on the beads. The cells were allowed to attach to the beads for 16 h at low speed then medium was added up to 500 ml and the stirrer speed increased to 30 rpm. After a growth phase of 48 to 72 h with daily aerating to allow gas exchange the beads were completely covered with cells and the medium was exchanged for production medium (Coons/DMEM, no fetal calf serum, penicillin and streptomycin sulfate at 100 μ g/ml each, 0.1% w/v Pluronic F68 and 5 mM NH₄Cl). The glucose concentration was monitored daily and the medium replaced, when glucose fell below 5 mM every 203 days. The harvested medium contained

approximately 2 mg. α-N-acetylglucosaminidase protein per dm³ of production medium.

EXAMPLE 8

Purification of recombinant α-N-acetylglucosaminidase

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Production medium was dialysed against 50 mM NaAc pH 5.5 and loaded onto a heparinagarose column equilibrated in the same buffer. After washing with NaAc buffer and NaAc/50 mM NaCl the column was eluted with 75 mM NaCl in NaAc buffer. The eluate was dialysed against 20 mM Tris/HCl pH 7.5, loaded onto a DEAE Scphacel column, washed with 25 mM NaCl in 20 mM Tris/HCl and then eluted with 50 and 75 mM NaCl in 20 mM Tris/HCl respectively.

SDS-PAGE of the two eluates showed two polypeptide bands associated with enzyme activity with apparent molecular weights of 79 and 89 kDa. The smaller α -N-acetylglucosaminidase was eluted predominantly in the 50 mM NaCl fraction whereas the 89 kDa α -N-acetylglucosaminidase polypeptide was enriched in the 75 mM NaCl fraction (Fig. 2).

The difference in apparent molecular weight of the recombinant α-N-acetylglucosaminidase polypeptides is due to the presence of additional carbohydrate side chains, since a digest with PNGase F, which cleaves off N-glycosylation groups, reduced both the 79 kDa and 89 kDa polypeptides to the polypeptide band having an apparent molecular weight of about 70 kDa (Fig. 2), which corresponds to the approximate molecular weight deduced from primary amino acid sequence data (SEO ID No:2).

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EXAMPLE 9

Characteristics of recombinant α-N-acetylglucosaminidase

No differences were observed between the enzyme activities of the 79 and 89 kDa recombinant α-N-acetylglucosaminidase polypeptides produced in CHO cells according to Example 7 and 8. With the fluorogenic N-acetylglucosamine α-linked to 4-

- 42 -

methylumbelliferone (4-MU) substrate, the enzyme has a pH-optimum of 4.6 with a k_M of 5.34 mM and a V_{max} of 3.97 x 10⁶ pmol/min/mg. Towards a ³H-labelled disaccharide substrate it should a pH-optimum of 4.1 with a k_M of 0.0166 mM and a V_{max} of 4.48 x 10⁴ pmol/min/mg.

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EXAMPLE 10

Mutational analysis of Sanfilippo B patients

Genomic DNA is isolated from cultivated skin fibroblasts of patients by extraction with

Phenol/Chloroform and used to amplify the eight exons and adjacent intronic sequences individually by PCR.

Primer sequences used in the amplification reaction are readily determined from the nucleotide sequences of the α-N-acetylglucosaminidase cDNA and genomic clones, set forth in SEQ ID No:1 or SEQ ID No:3. Amplification conditions are also readily determined without undue experimentation. Procedures for the design of PCR primers and amplification conditions are described in detail, for example, by McPherson *et al.* (1991). Differences in the primary sequence can be identified by separating the PCR products on a polyacrylamide gel under non-denaturing conditions (SSCP gels). Base changes, insertions and deletions will lead to a different band pattern compared with the wildtype in most of the cases, which can be visualised either by autoradiography of the gel after labelling the DNA during the PCR or by staining unlabelled DNA in the gel with silver. PCR products which show a different band pattern are sequences to identify the change. Other patient samples can be tested for mutations and polymorphism that were found by hybridisation with wildtype- and mutation-specific oligonucleotides (ASO).

Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of the steps, features, compositions and compounds referred to or indicated in this specification, individually or collectively, and any and all combinations of any two or

more of said steps or features.

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 Nature Genetics (submitted)

- 45 -

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: WOMEN'S AND CHILDREN'S HOSPITAL

(US ONLY: HOPWOOD JJ, WEBER B SCOTT HS, BLANCH LC, ANSON D)

- (ii) TITLE OF INVENTION: SYNTHETIC MAMMALIAN \(\alpha - N - ACETYLGLUCOSAMINIDASE \) AND GENETIC SEQUENCES ENCODING SAME
- (iii) NUMBER OF SEQUENCES: 6
- (iv) CORRESPONDENCE ADDRESS: (...
 - (A) ADDRESSEE: DAVIES COLLISON CAVE
 - (B) STREET: 1 LITTLE COLLINS STREET
 - (C) CITY: MELBOURNE
 - (D) STATE: VICTORIA
 - (E) COUNTRY: AUSTRALIA
 - (F) ZIP: 3000
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: AU PN6748/95
 - (B) FILING DATE: 23-NOV-1995
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: PCT INTERNATIONAL
 - (B) FILING DATE: 22-NOV-1996
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: HUGHES, DR E JOHN L
 - (C) REFERENCE/DOCKET NUMBER: MRO/EJH/JMC
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE:+61 3 9254 2777
 - (B) TELEFAX: +61 3 9254 2770

(2) INFORMATION FOR SEQ ID NO:1:

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		ACG														929
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	CTG Leu 550								1793
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	GCC Ala								1889
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	AGC Ser								1985
	GTC Val 630								2033
	CTG Leu								2081
	CAG Gln								2129
	TTC Phe								2177
	CAG Gln				-				2225
	CTC Leu 710								2273

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(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 743 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

(ix) FEATURE:

- (A) NAME/KEY: Potentially-glycosylated Asn site,
- (B) LOCATION: 261

(ix) FEATURE:

- (A) NAME/KEY: Potentially-glycosylated Asn site,
- (B) LOCATION: 272

(ix) FEATURE:

- (A) NAME/KEY: Potentially-glycosylated Asn site,
- (B) LOCATION: 435

(ix) FEATURE:

- (A) NAME/KEY: Potentially-glycosylated Asn site,
- (B) LOCATION: 503

(ix) FEATURE:

- (A) NAME/KEY: Potentially-glycosylated Asn site,
- (B) LOCATION: 513
- (ix) FEATURE:

- (A) NAME/KEY: Potentially-glycosylated Asn site,
- (B) LOCATION: 526

(ix) FEATURE:

- (A) NAME/KEY: Potentially-glycosylated Asn site,
- (B) LOCATION: 532
- Met Glu Ala Val Ala Val Ala Ala Ala Val Gly Val Leu Leu Leu Ala 1 5 10 15
- Arg Ala Leu Val Ala Arg Leu Leu Ġly Pro Gly Pro Ala Ala Asp Phe
 35 40 45
- Ser Val Ser Val Glu Arg Ala Leu Ala Ala Lys Pro Gly Leu Asp Thr 50 55 60
- Tyr Ser Leu Gly Gly Gly Ala Ala Arg Val Arg Val Arg Gly Ser
 65 70 75 80
- Thr Gly Val Ala Ala Ala Ala Gly Leu His Arg Tyr Leu Arg Asp Phe 85 90 95
- Cys Gly Cys His Val Ala Trp Ser Gly Ser Gln Leu Arg Leu Pro Arg 100 105 110
- Pro Leu Pro Ala Val Pro Gly Glu Leu Thr Glu Ala Thr Pro Asn Arg
- Tyr Arg Tyr Tyr Gln Asn Val Cys Thr Gln Ser Tyr Ser Phe Val Trp 130 135 140
- Trp Asp Trp Ala Arg Trp Glu Arg Glu Ile Asp Trp Met Ala Leu Asn 145 150 155 160
- Gly Ile Asn Leu Ala Leu Ala Trp Ser Gly Gln Glu Ala Ile Trp Gln 165 170 175
- Arg Val Tyr Leu Ala Leu Gly Leu Thr Gln Ala Glu Ile Asn Glu Phe 180 185 190
- Phe Thr Gly Pro Ala Phe Leu Ala Trp Gly Arg Met Gly Asn Leu His
 195 200 205
- Thr Trp Asp Gly Pro Leu Pro Pro Ser Trp His Ile Lys Gln Leu Tyr 210 215 220
- Leu Gln His Arg Val Leu Asp Gln Met Arg Ser Phe Gly Met Thr Pro 225 230 235 240

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Val	Leu	Pro	Ala	Phe 245	Ala	Gly	His	Val	Pro 250		Ala	Val	Thr	Arg 255	Val
Phe	Pro	Gln	Val 260	Asn	Val	Thr	Lys	Met 265	Gly	Ser	Trp	Gly	His 270	Phe	Asn
Суэ	Ser	Tyr 275	Ser	Сув	Ser	Phe	Leu 280	Leu	Ala	Pro	Glu	Авр 285	Pro	Ile	Phe
Pro	Ile 290	Ile	Gly	Ser	Leu	Phe 295	Leu	Arg	Glu	Leu	Ile 300	Lys	Glu	Phe	Gly
Thr 305	qaA	His	Ile	Tyr	Gly 310	Ala	Авр	Thr	Phe	Asn 315	Glu	Met	Gln	Pro	Pro 320
Ser	Ser	Glu	Pro	Ser 325	Tyr	Leu	Ala	Ala	Ala 330	Thr	Thr	Ala	Val	Tyr 335	Glu
Ala	Met	Thr	Ala 340	Val	Asp	Thr	Glu	Ala 345	Val	Trp	Leu	Leu	Gln 350	Gly	Trp
Leu	Phe	Gln 355	His	Gln	Pro	Gln	Phe 360	Trp	Gly	Pro	Ala	Gln 365	Ile	Arg	Ala
Val	Leu 370	Gly	Ala	Val	Pro	Arg 375	Gly	Arg	Leu	Leu	Val 380	Leu	Asp	Leu	Phe
Ala 385	Glu	Ser	Gln	Pro	Val 390	Tyr	Thr	Arg	Thr	Ala 395	Ser	Phe	Gln	Gly	Gln 400
Pro	Phe	Ile	Trp	Сув 405	Met	Leu	His	Asn	Phe 410	Gly	Gly	Aвп	His	Gly 415	Leu
Phe	Gly	Ala	Leu 420	Glu	Ala	Val	Asn	Gly 425	Gly	Pro	Glu	Ala	Ala 430	Arg	Leu
Phe	Pro	Asn 435	Ser	Thr	Met	Val	Gly 440	Thr	Gly	Met	Ala	Pro 445	Glu	Gly	Ile
Ser	Gln 450	Asn	Glu	Val	Val	Tyr 4 55	Ser	Leu	Met	Ala	Glu 460	Leu	Gly	Trp	Arg
Lys 465	Asp	Pro	Val	Pro	Asp 470	Leu	Ala	Ala	Trp	Val 475	Thr	Ser	Phe	Ala	Ala 480

Arg Arg Tyr Gly Val Ser His Pro Asp-Ala Gly Ala Ala Trp Arg-Leu

Leu Leu Arg Ser Val Tyr Asn Cys Ser Gly Glu Ala Cys Arg Gly His

Asn Arg Ser Pro Leu Val Arg Arg Pro Ser Leu Gln Met Asn Thr Ser

505

485

500

- 53 -

Ile Trp Tyr Asn Arg Ser Asp Val Phe Glu Ala Trp Arg Leu Leu Leu 530 535 540

Thr Ser Ala Pro Ser Leu Ala Thr Ser Pro Ala Phe Arg Tyr Asp Leu 545 550 555 560

Leu Asp Leu Thr Arg Gln Ala Val Gln Glu Leu Val Ser Leu Tyr Tyr
565 570 575

Glu Glu Ala Arg Ser Ala Tyr Leu Ser Lys Glu Leu Ala Ser Leu Leu 580 585 590

Arg Ala Gly Gly Val Leu Ala Tyr Glu Leu Leu Pro Ala Leu Asp Glu
505 600 605

Val Leu Ala Ser Asp Ser Arg Phe Leu Leu Gly Ser Trp Leu Glu Gln 610 615 620

Ala Arg Ala Ala Ala Val Ser Glu Ala Glu Ala Asp Phe Tyr Glu Gln 625 630 635 640

Asn Ser Arg Tyr Gln Leu Thr Leu Trp Gly Pro Glu Gly Asn Ile Leu 645 650 655

Asp Tyr Ala Asn Lys Gln Leu Ala Gly Leu Val Ala Asn Tyr Tyr Thr
660 665 670

Pro Arg Trp Arg Leu Phe Leu Glu Ala Leu Val Asp Ser Val Ala Gln 675 680 685

Gly Ile Pro Phe Gln Gln His Gln Phe Asp Lys Asn Val Phe Gln Leu 690 695 700

Glu Gln Ala Phe Val Leu Ser Lys Gln Arg Tyr Pro Ser Gln Pro Arg 705 710 715 720

Gly Asp Thr Val Asp Leu Ala Lys Lys Ile Phe Leu Lys Tyr Tyr Pro
725 730 735

Gly Trp Val Ala Gly Ser Trp 740

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 10380 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)

(vi) ORIGINA	L SOURCE:
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(A) ORGANISM: Homo sapiens

(viii) POSITION IN GENOME:

(A) CHROMOSOME/SEGMENT: Chromosome 17

(ix) FEATURE:

(A) NAME/KEY: exon 1

(B) LOCATION: 990..1372

(ix) FEATURE:

(A) NAME/KEY: exon 2

(B) LOCATION: 2115..2262

(ix) FEATURE:

(A) NAME/KEY: exon 3

(B) LOCATION: 3056..3202

(ix) FEATURE:

(A) NAME/KEY: exon 4

(B) LOCATION: 3387..3472

(ix) FEATURE:

(A) NAME/KEY: exon 5

(B) LOCATION: 5667..5923

(ix) FEATURE:

(A) NAME/KEY: exon 6

(B) LOCATION: 7745..8955

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ATAATGAGCA GTGAGGACGA TCAGAGGTCA CCTTCCTGTC TTGGTTTTGG CAGGTTTTGA CCAGTTTCTT TGCTGCATTC TGTTTTATCA GCGGGGTCTT GTGACCTTTT ATCTTGTGCT 120 GACCTCCTGT CTCATCCTGT GACGAAGGCC TAACCTCCTG GGAATTCAGC CCAGCAGGTC 180 TCTGCCTCAT TTTACCCAGC CCCTGTTCAA GATGGAGTCG CTCTGGTTGG AAACTTCTGA 240 CAAAATGACA GCTCCTGTTA TGTTGCTGCT GCTGCCGCCA ATGGACAGCC TTTAACGTGC 300 CCGCCAGCCC TGCTCCACCG CCGGCCTGGG CTCACATGGC CCCATCCCTC CTCGAACCTC 360 CTAGCCTGTT AGTTACTCAA ATCTGCAAGC TCTCTGCCTT CTCAGGGCCT TCAATAAATG 420 CATTTCTTCT GTCTGGAAGG CTCTTCCTTT CCCTCTTCTA GCCAATTCCT ATTCATCCCT GAGTTTCAGA TTAAAAGTCA CTTCCTTTGG AAACCTTACT TCGCTACTTC GCTACTTACT GCACTACTTC GCAGCATCAC AACTATGATG GAAATCCTTA CTTACGTTAA ATATCTGGTT 600

660	AGTAGGGAAG	CTCGTTCATC	GGACCGTCTT	GGGACGGTAG	CTCCCTGACG	TCTAGGTCAC
720	GATGGTTGGA	GTATTTATTA	CTCCAAATGT	ACAAAATAAA	AGTGCCTGAT	TAGCTATGGC
780	GGGCCAGCGG	CGCTCTGTGA	TACCCGAAGG	AAGCGCGTTT	TTGCGTGTGA	TGGAAGTTAT
840	CACGAGACGC	CGCGTGCGGT	CGCTCCCCAC	CGGGGTCACA	GCCCTGGAGC	GTCCCCTTCG
900	CCGGGCTTAG	GGCCCTGAGC	CGCGACTCCT	ACCCGGAAGC	GTATCCTGGT	CCCCAAGGGA
960	CCACCCCCTG	GCGGGCCGCC	CTGATTGGAC	GAGCCGGCAG	CACGTGGCCG	CCTTCGGGTC
1020	GCGGCGGTGG	GGCGGTGGCC	TGGAGGCGGT	ACTGAGACCA	GACCCGCAGG	GCCGTCGCGG
1080	GAGGCGGCGG	·CGAGGCCCGG	CGGCAGGCGA	GCCGGGGGCG.	CCTGGCCGGG	GGTCCTTCT
1140	TTCTCCGTGT	CGCGGCCGAC	GGCCAGGCCC	CGGCTGCTGG	GCTCGTGGCC	CCGTGCGGGC
1200	GGCGGCGGCG	CTACAGCCTG	GCTTGGACAC	GCCAAGCCGG	CGCTCTGGCT	CGGTGGAGCG
1260	GGGCTGCACC	GGCCGCGCG	CGGCGTGGC	CGCGGCTCCA	CGTGCGGGTG	gcgcgcgcg
1320	CTGCGCCTGC	CGGCTCTCAG	TGGCCTGGTC	GGCTGCCACG	CGACTTCTGT	CTACCTGCG
1380	AGGTACCGCC	CACGCCCAAC	TGACCGAGGC	CCGGGGGAGC	GCCAGCCGTG	CGCGGCCACT
1440	TGCCACCCAA	CCGGAGCCGC	TTACCCCCTC	CCCGAGGCGC	CCCGCGTCCG	CCGAAGCTTC
1500	TCCAGCAGCT	CTGCGCCGCC	GAAGGCCCAG	GCGCTGGCCG	TGAGCGGGGA	ATCGGGAGGC
1560	CTGAAAAACG	GCTGGCCCAC	CAGAGCCTCG	CTCTGCCTTT	GAGCCAGCCA	STGTGGCCTT
1620	GGGCATAGAA	GCACGATGAT	GTGAGGATTT	CAGTGTTATT	GCCTACCGTG	Gaaagagac
1680	CCTTCTCTTT	TCCTCCCCCA	TTTCTTGCCT	ATGAGTGAAT	ACAATTGGTG	TTTGTGGTGC
1740	TGTTCCAGCG	TACAGGCCTG	TCCCCCACCC	GACGCCTCCA	ACTGAGGAAG	BAACCTGCGG
1800	TCTGTTAGAC	CCCCTGCCCA	CAGCTGTCCT	TGTGTTCACA	TATGGAGTGA	CCTGCCACAC
1860	CTCTGGCAGT	AGAGGAGGG	AACACCGTGC	GTTCCAGGAA	GGGATTCCCC	TGTGGGGGCA
1920	CCTGGGAGAG	GCTAGAGGGC	TACCCGCCAG	GCCACCCAAA	AGTGGAATAT	STGGCATGAA
1980	CCACCTGGGG	GGTCTCAGCT	CCCGGTACCT	GAAGCCCAGC	GAGTGCCTCA	rgcaggggac
2040	CTCTAGGTGG	стсссстстс	TTTGGAGCCC	AAGGGCCGAG	TGTGCAGCAG	GGGTCCCAG
2100	CCCTGCTCAT	TGGGATGCGC	CCCTCCAGGG	GGGCCGTGGA	ATTTGTTCCA	GGATGGGGG
2160	CTCCTTCGTG	CGCAAAGCTA	AATGTGTGCA	CTATTACCAG	GCAGGTACCG	SACACTGCCC
	modormarr.	magagama* *	> m> a> amaa>	GGNGGGNGNG	0000000000	CCTCCCA

WO 97/19177

CTGGCACTGG CCTGGAGCGG CCAGGAGGCC ATCTGGCAGC GGGTGCGTGC CCACTGTCCC 2280 TTCCCCACCC TCCTCTATGG CGGGAGCCAC CGTAGGTGTT TTCACCCGCC CCCCAGCATG 2340 GGCGCAGTGT CTCTCTAG AAGTGCTTTC AGCGTGCACA GTGGCTTGGG CCTCCTAAAA 2400 ACTGAGGCTT CCGGCCGGGC GCGGTGGCTC ACGCCTGTCA TCCCAGCACT TCGGGAGGCC 2460 TAGGCGGCC GATCAGGAGT TCAGGAGATC GAGACCATCC TGGCCAACAT TGTGAAACCC 2520 CGTCTCTACT AAAATACAAA GAAATAGCAA CCTGGGCAAC AGAGCGAGAC TCTGTCTAAA 2580 AAAAAAAAA AAAAAAACTG AGGCTTCCAG TTTGAGGAGT GGGGCTCCTT CCCCCATCTC 2640 CCCTATGCAG CCAATCACCT GGTCCCTTGG ATCCAACTCA TGGGCAGCTC TAGATCTGCC 2700 TCCCTGGAAG CTTCTGTGCT GCAATGGCTG CTCCAGGCTC TGCTTAAGCT CTTCACACAG 2760 TTGCCCTGCC CTTCCATCTG GCACTCTTGC TCCATGAAGC CTTCTAAGGC CTTCCTGTTG 2820 GGGGAAAGCC CCTTTGTGCC CCATCTCCTC ACCCATGCGA CAAAGGCAAC ACAGTGAACT 2880 CACCTACTCA CAGGTCTCTT TCCTCTGGGC TGTGGGCTCC TTGATGGCAG CGTTCGGATT 2940 TTGTCTCAGT AGCCCTAGCA CCCAGCACAA AGAAGCAATG AGTGAATGGT TGTTGAATGA 3000 ATGAATGAAT GAATGAAGAT GAATATATTT CTATGTGTGG GCCCTTCTTC CTCAGGTGTA 3060 CCTGGCCTTG GGCCTGACCC AGGCAGAGAT CAATGAGTTC TTTACTGGTC CTGCCTTCCT 3120 GGCCTGGGGG CGAATGGGCA ACCTGCACAC CTGGGATGGC CCCCTGCCCC CCTCCTGGCA 3180 CATCAAGCAG CTTTACCTGC AGGTAAAAGG ATGGAAAAGG GAAGGGGCAG AATCGGTGAT AGATGGTCAT GGGCCCAGGA AGGGTGGTAT TAGGCCGGCC CCAGGGCTCT TAACTGAGGC 3300 GGGGGGCTGC GTGTATCCTG GGAGATGAGG GCCTTCTCAT AGGACAGCAG TGGCCATGCT 3360 CACCACCCTT CCTTCTGTTC CTCCAGCACC GGGTCCTGGA CCAGATGCGC TCCTTCGGCA 3420 TGACCCCAGT GCTGCCTGCA TTCGCGGGGC ATGTTCCCGA GGCTGTCACC AGGTGAGGTT 3480 CCGCTCACCC CCTCCACTTA GCTCAGAGAG GGAATTTTAT TCCCTTCTAG AACATGACTT 3540 AAAAACTTAA GCTCTGGGCC GGGCGCAGTG GCTCACGCCT GTAATCCCAG CACTTTGGGA GGCCGAGTTG GGCGGATCAC CTGAGGTCAG GAGTTCGAGA CCAGCCTGGC CAACATGGTG 3660 AAACCCTGTC TCTACTAAAA ATATAAAAAT TAGCTGGGCA TGGTGGCACG CGCCTGTAAT 3720 CCCATCTACT TAGGAGGCTG AGACAGGAGA ATTGCTTAAA CCTGGGAGGC AGACGTTGCA 3780 GTGAGTCAAG ATCACGCCAT TGCACTCCAG CCTGGGTGAC GAGCGAAACT CTGTCTCAAA 3840

WO 97/19177

CAMACAMAC	A AGCICIGGAC	. GINGGCCIGC	GIIIGAIIIC	. IGACICIGCI	ACIAAIIAGC	390
TGTGTGACT	T CGGGCAGATO	ACATGACTG	TCTGTGCCTC	: AGTTTCCTTA	CTTGTAAAAT	396
GGGATCTCT.	A CCCACTTCGC	TGTAGGGTT	GTAATTATCT	CTCGATCTAT	CTGTGACTTT	402
GCACAGAGT	G CTAGCAAATG	GCAGCCCTTG	GGAGTGGCAG	CAGGGGTGCT	CCAGTGTCCC	408
TTGTCCCTC	с теттестете	TGCTTCCCAG	CCATCCTCTC	ACATGTGGTT	GGGAAAAGTC	414
TTCAAGGCT	C ACCTGAGACC	: TCCCCTCCTT	CAGGAAGCCT	TGCTAGTGCC	CCGCATGACC	420
TCCTTTGCA	C CTGCTAATGT	CTGGCTCCCA	TACTCTCGTA	GGACTTAATG	CATGCCAGTG	426
GCCTCCCTG	C CCGCCTCTTT	GCCCCCATCA	CCAGGTGGCA	GGAAACTCAC	TCATTCATTC	432
AATAAACTT	GTCCAGCTGT	CTGAGGCTGC	CAGAACTGGC	TGTGCTGGGT	CCTGGGAGGC	438
GGCAAGAAA	G GTGCCCAAGG	GCTTACCCCT	GATAGGAGAG	ATATGTTGGC	TGAAGGATAC	444
aatgtgggg <i>i</i>	A CAAGGACAGG	AATATATGTG	GGTTCCGCTC	TCCTCTGCCG	GGAGAGAGGG	450
GCAGGAAGG	G CTCAGGGCAG	AGCCCAGCCT	TGAAAAATGA	GTGTTGCTTG	GACGGACGCT	456
rggctaatg(TTGTAATCCT	AGCGTTTTGG	GAGGCTGAGG	CGTATGGATC	ACCTGCGGTC	4620
AGGAGTTAAA	GACCAGCCTG	GCCAACATGG	CGAAACCCCA	TCTCTACTAA	AAGTACAAAA	4686
ATTAGCCAGG	GCTGGTGGCG	GGCTCCTGTA	ATCCCAGCTA	CTCGGTAGGC	TGAGGCATGA	4740
GAATCTCTTG	AAGCCAGGGG	CCAGAGACTG	CAGTGAGCCG	AGATCACACC	ACTTCACTCC	4800
AGCCTGGGTC	ACAGAGTGAG	ACTCCGTCTC	AAAAAAAA	AAAAAAAAG	GAAAGAAAAT	4860
TAAACACCTC	ATGTTCTCAC	TCATAGTGGG	AGTTGAACAA	TGAGAACAAC	ATGGACACAG	4920
BAAGGGGAAC	* ATCACACACC	GGGCCTTTC	GCGGTGTGGG	GGTCAAGGGG	AGGAGTAGCA	4980
TGGGACAGA	TACTTAATGC	ATGCGGGGCT	GAAAACCTAG	ATGATGGGTT	GATGGGTGCA	5040
CAAACCACC	ATGGCACATG	TATACCTATG	CAACAAACCT	GCATGTTCTG	CACAGAACTG	5100
actgaaagi	` ATAATTAAAA	ааааааааа	AAGCTGGGTG	CGGTGGCCCA	CACCTGTAAT	5160
CCAGCACTI	TGGGAGGCCG	AGACGGGCGG	ATCACAAGGT	CAGCAGATCG	AGACCATCCT	5220
IGCTAACACA	GTGAAACTCA	GTCTCTACTA	AAAATACAAA	AAATTAGCCG	ggtgtgg t gg	5280
GGGCACCTG	TAGTCCCAGC	TACTAGGGAG	GCTGAGGCAG	GAGAATGGCA	TGAACCTGGG	5340
GGCAGAGCT	TGCAGTGAGC	TGAGAATGCG	CCACTGCACT	CCAGCCTGGG	GGACAGAGTG	5400
GACTCTGCC	ТСАЛАЛАЛА	DAAAAAAAAG	AAAGAAAAG	GAGCGTTGCT	TGTTTCAGGC	5460

CACAGGAAGG	G GGAGAGATAC	G TGAAAGTTT	r TCAGAGAAGG	TGGCCAGGGA	AGGAGAAGAA	552
AGGACTGTAG	GCAGAGAGC	A TAGCCTGTAG	CAAAGCCATAG	AGGCAAGAGA	AACCAGGAGC	558
TGTAGAGAAG	TTGGCAAGG	TGTTGAACAC	TATGGTGAAC	ACTATGGCGG	CTTCCATGAA	564
ATATCTGAGC	TTTTGCTCCC	CACTAGGGTG	TTCCCTCAGG	TCAATGTCAC	GAAGATGGGC	5700
AGTTGGGGCC	: ACTTTAACTG	TTCCTACTCC	TGCTCCTTCC	TTCTGGCTCC	GGAAGACCCC	5760
ATATTCCCCA	TCATCGGGAG	CCTCTTCCTG	CGAGAGCTGA	TCAAAGAGTT	TGGCACAGAC	5820
CACATCTATG	GGGCCGACAC	TTTCAATGAG	ATGCAGCCAC	CTTCCTCAGA	GCCCTCCTAC	5880
CTTGCCGCAG	CCACCACTGC	CGTCTATGAG	GCCATGACTG	CAGGTACAGT	GCCTGGGTGG	5940
ggtgggagag	CCCCCAGAC	CCTCAAAAAG	AAGGGAGTAG	CAGATGTCAG	TAGGGGTAGG	6000
CAGAGGGACT	GGAATAATGC	CTCGCCATAA	CACACAGTAC	TTTATAGTTT	ACCAAGCACG	6060
TGTACACATG	CGTTGTCTCA	GTGAATCCCA	CTGTGGTTGA	GAGGTGAGCT	CTGGAAGCCA	6120
ACAACCTGGG	TCACACCTCG	CGCTCCTATT	TCCTGGCCGT	GTGACTTATG	ACTCATGACC	6180
TCCTTCCCAG	TGTCTCGTTT	GCTTTTCCTG	TAAACTGGGA	CTACCTCATA	GGTAGAATAA	6240
CGCCTGGCCC	AGAGCAAAGG	CCACTAAGAG	CTAGCTATGA	ACAAGGATTT	TGTTTCATCT	6300
CTGCGTGGTT	GCTGAAGTAG	GCACTGCAGG	CAGGAGGTGA	GTGGATGTGC	CTAAAGGCAC	6360
TAAGTGCGCA	TCCTGCTACA	AAACTGTGAA	GCCAGGGCTC	CTTCCTGCCA	CTTAAAGGAG	6420
GAGTGGAGCA	GAGGGCGCCC	AAGTCAGGAA	TGACTTAGTG	GAGAGGCGTC	TGTGTTGGCC	6480
AGGAAGGGAA	CAGATCAGCT	CAGCCTTTCT	TGAGCAGTAC	TGCTCCAAGT	GTGACCCAAA	6540
ACCAGCAGCA	GCAGCAGCAG	CAGCCCGAGC	TGTGAGATGG	CAAATTCTCA	GGCCCTACCC	6600
AAGACCTGAA	GGAGAAGCTA	CATTTTTTT	TTTTTTGAGA	CAGATTTCAC	TCTGTTGCTG	6660
AGGCTGGAGC	ACAGTGGCAC	AATCTCATCT	CACTGCAACC	TTCGTCTCCT	AGGTTCAAGC	6720
BATTCTCCTG	CCTCAGCCTC	CCGAGTAGCT	GGGACTATAG	GCACCCGCCA	CCACGCCCGG	6780
CAATTTTTGT	TTGTTTTGAG	ATAGAGTCTC	GCTCTGTCAC	CCAGGCTGGA	GTGCAGTGGC	6840
ACGATCTCAG	TTCACTGCAA	CCTCTGCTTC	CTGAGTTCAA	GCGATTCTCC	TGCCTCAGCC	6900
CCTGAGTAG	CTGGGATTAC	AGGCGCCCC	CAACCACACT	CGGCTAATTT	TTGTATTTT	6960
GTAGAGACG	GGGTTTCGCT	ATGTAGGTCA	AGCTGGTTTC	AAACTCCTGA	CCTCAAATGA	7020
TCGCCCACT	TCAGCCTCCC	AAAGTGCTGG	GATTACAGGT	GTGAGCCACC	TTGCCTGGCC	7080

A	ATTTTTGTA	TTTTTAGTAG	AAACAGGTTT	CACCATGGTG	GCCAGACTGG	TCTCAAACTC	7140
C.	TGACCTCAG	GTGAACTGCC	CACCTCAGCC	TCCCAAAGTA	CTGGTATTAC	AGGCGTGATC	7200
C	ACTGCGACT	GGCCTTGATT	TTGTTTTTGA	GACAGAATCT	TACTCTGTCG	CCCAGACTGG	7260
A	GTGCAGTGG	CACAATCTCA	GCTCACTGCA	ACTTCTGCCT	CATGGGTTCA	AGTGATTCTT	7320
G'	TGCCTCTAC	CTCCCGAGTA	GCCGGGATTA	CAGGCACCTG	CCATTACGCT	AGGCTAATTT	7380
T.	IGTATTTT	AGTATAGACA	GGGTTTCCCC	ACATTGGCCA	GGCTGGTCTG	GAACTCCTGG	7440
G	CTCAAGTGA	TCCACCTGCT	TCAGCCCCTC	AGAGTACTGG	GATTATAGGT	GTGGGCCACC	7500
A	CGCCCATTC	AGAAACCTCC	ATGTTTTAAG	GAGCCCTCTG	GGTAACTCTC	ATGTTCACCC	7560
A	agctgctga	ACCCTGTCCT	GGAGTTTTCA	GAGGGACGCG	TATGTGCCAC	AGAGCGTCCC	7620
G	CTGGTGGGG	GTCATGGGAA	GCCATGACCT	GGGATAGACA	GTCGTCTGTA	GAGTGGGGTG	7680
A.J	ACATTCCCT	GGGCCCTCTG	TTTCATCACT	ССТСТТСТСТ	GTTCCCCCTA	CCTCCTGTCC	7740
AC	CAGTGGATA	CTGAGGCTGT	GTGGCTGCTC	CAAGGCTGGC	TCTTCCAGCA	CCAGCCGCAG	7800
T	CTGGGGC	CCGCCCAGAT	CAGGGCTGTG	CTGGGAGCTG	TGCCCCGTGG	ссвсстсств	7860
G7	TCTGGACC	TGTTTGCTGA	GAGCCAGCCT	GTGTATACCC	GCACTGCCTC	CTTCCAGGGC	7920
CA	GCCCTTCA	TCTGGTGCAT	GCTGCACAAC	TTTGGGGGAA	ACCATGGTCT	TTTTGGAGCC	7980
CI	AGAGGCTG	TGAACGGAGG	CCCAGAAGCT	GCCCGCCTCT	TCCCCAACTC	CACCATGGTA	8040
GG	CACGGGCA	TGGCCCCCGA	GGGCATCAGC	CAGAACGAAG	TGGTCTATTC	CCTCATGGCT	8100
GA	CTGGGCT	ggcgaaagga	CCCAGTGCCA	GATTTGGCAG	CCTGGGTGAC	CAGCTTTGCC	8160
GC	CCGGCGGT	ATGGGGTCTC	CCACCCGGAC	GCAGGGGCAG	CGTGGAGGCT	ACTGCTCCGG	8220
AG	TGTGTACA	ACTGCTCCGG	GGAGGCCTGC	AGGGGCCACA	ATCGTAGCCC	GCTGGTCAGG	8280
· CG	GCCGTCCC	TACAGATGAA	TACCAGCATC	TGGTACAACC	GATCTGATGT	GTTTGAGGCC	8340
TG	GCGGCTGC	TGCTCACATC	TGCTCCCTCC	CTGGCCACCA	GCCCGCCTT	CCGCTACGAC	8400
CT	GCTGGACC	TCACTCGGCA	GGCAGTGCAG	GAGCTGGTCA	GCTTGTACTA	TGAGGAGGCA	8460
AG	AAGCGCCT	ACCTGAGCAA	GGAGCTGGCC	TCCCTGTTGA	GGGCTGGAGG	CGTCCTGGCC	8520
TA	TGAGCTGC	TGCCGGCACT	GGACGAGGTG	CTGGCTAGTG	ACAGÇCGCTT	CTTGCTGGGC	8580
AG	CTGGCTAG	AGCAGGCCCG	AGCAGCGGCA	GTCAGTGAGG	CCGAGGCCGA-	TTTCTACGAG	8640
CA	GAACAGCC	GCTACCAGCT	GACCTTGTGG	GGGCCAGAAG	GCAACATCCT	GGACTATGCC	8700

PCT/AU96/00747

AACAAGCAGC	TGGCGGGGTT	GGTGGCCAAC	TACTACACCC	CTCGCTGGCG	GCTTTTCCTG	876
GAGGCGCTGG	TTGACAGTGT	GGCCCAGGGC	ATCCCTTTCC	AACAGCACCA	GTTTGACAAA	8820
AATGTCTTCC	AACTGGAGCA	GGCCTTCGTT	CTCAGCAAGC	AGAGGTACCC	CAGCCAGCCG	888
CGAGGAGACA	CTGTGGACCT	GGCCAAGAAG	ATCTTCCTCA	AATATTACCC	CGGCTGGGTG	8940
GCCGGCTCTT	GGTGATAGAT	TCGCCACCAC	TGGGCCTTGT	TTTCCGCTAA	TTCCAGGGCA	9000
GATTCCAGGG	CCCAGAGCTG	GACAGACATC	ACAGGATAAC	CCAGGCCTGG	GAGGAGGCCC	9060
CACGGCCTGC	TGGTGGGGTC	TGACCTGGGG	GGATTGGAGG	GAAATGACCT	GCCCTCCACC	9120
ACCACCCAAA	GTGTGGGATT	AAAGTACTGT	TTTCTTTCCA	CTTAAACTGA	TGAGTCCCCT	9,180
GGGTCTGTCA	AAATGAGAAG	GTCACTGCTG	CCACGCTTGG	GAGGACTCAG	GGCTATAGCA	9240
TGGCCCTGGG	GTGGGACCTG	TTCTCCCATC	CCTTGCCTCA	CGTCCCTGTT	TTTGTTTGTT	9300
TGTTTGTTTG	TGACGGAGCC	TTGGTCTGTT	GCCCAGGCTT	GAGTACAATG	GCACAGTCTC	9360
GGCTCACTGC	AACCTCCGCC	TCCTGGGTTC	AAGCAATTCT	TGTGCCTCAG	CCTCCCCGGT	9420
AGCTGGGACT	ATAGGCATGC	ACCACCACAC	CAGGCTAATT	ттттттттс	CAAGATGGAG	9480
TCTTGCTCTG	TCGCCCAGGT	TGGAGTTTAG	TGGCACCATA	TTGGTTTACT	GCAACCTCTG	9540
CCTCCCGGGT	TCAAGCAATT	CTCCTGCCTC	AGTCTACCAG	GGAGTTAGGA	CTACGGGCCT	9600
GTGCCATCAC	GCCTGGCTAA	TTTTTGTATT	TTTCATAGAG	ATAAGGTTTC	ACCATGTTGG	9660
CCAGGCTGGT	CTTTAACTCC	TGAACTCAAG	TGATCCACCT	GCCTCGGCCT	TCCAAAGTGC	9720
TGGGATTACA	GGAGTGAGCC	ACCGTGCCCG	GCCATGTCTC	TCTTTTTAAC	ACTAATGTTA	9780
CCCTGACCTT	TGAACGTAGA	ATGCCCTTCT	GTTGCAGGAA	AACCTCTTTT	CAAACCATGT	9840
TTGTCCTTTG	CTGGCATGCC	ACAGCAACAG	TCACCAACAC	AGAAGACTTC	TGTGACCAAA	9900
TATTTGGAGG	ATTTTCCCCA	CACACACCAA	GCAGCAGACA	TCAGCTGGGT	GTCCTCCAAT	9960
TCAGTTCCAA	TGTAATCAAC	CAGAGACAGC	ATCAGATCCC	ACAGGGTTAG	GGTGCAGATC	10020
CATGAGACCA	CCCCCTCCTT	CCCAACGGTT	ACAAGTCCTG	ATCCCTGGAA	CTTCTGACTA	10080
ACTGGCTTCA	AGTTGGAGTT	CCCATGACCC	CCTTCCCCTC	TTTGGAGTCA	ACTCATTTGC	10140
GACAGTGACC	CACGAAACAC	AGGGAAACCC	TTATTATGTT	TATTGCTTTA	TTACAGAGGA	10200
AAAAAATTTT	TTTCTTTCTT	TTTTGAGACA	GGGTCTCACT	CTGTCATCCA	GAATGACTGC	10260
AGTGGCAGGA	TCTGGCTCCG	TCACCCAGGC	TGGAGTGCAG	TGGCATGATC	TCGGCTCACT	10320

PCT/AU96/00747

-61-

ACAGCCTCCA TCCCCCCCAA ACCCCACGCC TCAGCGCCCC ACCCCGCAAG TGGCTGGGAC 10380

- (2) INFORMATION FOR SEQ ID NO:4:
 - (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: peptide
- (iii) HYPOTHETICAL: NO
 - (v) FRAGMENT TYPE: N-terminal
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Homo sapiens
- (ix) FEATURE:
 - (A) NAME/KEY: Modified-site
 - (B) LOCATION: 10
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Asp Glu Ala Arg Glu Ala Ala Ala Val Arg Ala Leu Val Ala Arg Leu 1 5 10 15

Leu Gly Pro Gly

20

- (2) INFORMATION FOR SEQ ID NO:5:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 18 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: NO
 - (v) FRAGMENT TYPE: N-terminal
 - (vi) ORIGINAL SOURCE:

(A) ORGANISM: Homo sapiens

(ix) FEATURE:

- (A) NAME/KEY: Modified-site, glycosylated or phosphorylated, wherein Xaa may be any amino acid residue, preferably Arg.
- (B) LOCATION: 16
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Lys Pro Gly Leu Asp Thr Tyr Ser Leu Gly Gly Gly Gly Ala Ala Xaa Val

Arg

- (2) INFORMATION FOR SEQ ID NO:6:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: peptide
 - (iii) HYPOTHETICAL: NO
 - (v) FRAGMENT TYPE: internal
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Homo sapiens
 - (ix) FEATURE:
 - (A) NAME/KEY: Modified-site, glycosylated or phosphorylated, wherein Xaa may be any amino acid residue, preferably Ala
 - (B) LOCATION: 12
 - (ix) FEATURE:
 - (A) NAME/KEY: Modified-site, glycosylated or phosphorylated, wherein Xaa may be any amino acid residue, preferably Ser
 - (B) LOCATION: 14
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Trp Arg Leu Leu Thr Ser Ala Pro Ser Leu Xaa Thr Xaa Pro 1 5 10 15

CLAIMS:

- 1. An isolated nucleic acid molecule comprising a sequence of nucleotides which encodes or is complementary to a sequence which encodes a mammalian α -N-acetylglucosaminidase or fragment or derivative thereof.
- 2. The isolated nucleic acid molecule according to claim 1 wherein the nucleotides are deoxyribonucleotides.
- 3. The isolated nucleic acid molecule according to claim 2 wherein said molecule is cDNA.
- 4. The isolated nucleic acid molecule according to claim 2 wherein said molecule is a genomic DNA molecule.
- 5. The isolated nucleic acid according to any one of claims 1 to 4 wherein the mammal is a human.
- 6. The isolated nucleic acid according to claim 5 wherein the α -N-acetylglucosaminidase is of liver, kidney or placenta origin.
- 7. The isolated nucleic acid molecule according to any one of claims 1 to 6 having a nucleotide sequence substantially as set forth in SEQ ID NO:1 or complementary thereto or having at least 40% similarity to all or part thereof.
- 8. The isolated nucleic acid molecule according to any one of claims 1 to 6 having a nucleotide sequence substantially as set forth in SEQ ID No:3 or complementary thereto or having at least 40% similarity to all or part thereof.
- 9. The isolated nucleic acid molecule according to claim 7 or 8 wherein the percentage similarity is at least 60%.

- 10. The isolated nucleic acid molecule according to claim 9 wherein the percentage homology is at least 80%.
- 11. The isolated nucleic acid molecule according to any one of claims 1 to 10 wherein the α-N-acetylglucosaminidase or fragment or derivative thereof encoded by said molecule comprises an amino acid sequence substantially identical to SEQ ID No:2 or is at least 40% similar to all or a part thereof.
- 12. The isolated nucleic acid molecule according to claim 11 wherein the percentage similarity to SEQ ID No:2 is at least 60%.
- 13. The isolated nucleic acid molecule according to claim 12 wherein the percentage similarity to SEQ ID No:2 is at least 80%.
- 14. The isolated nucleic acid molecule according to any one of claims 1 or 13 wherein said molecule is carried by a vector capable of replication in a eukaryotic cell and/or a prokaryotic cell.
- 15. The isolated nucleic acid molecule according to claim 14 wherein the vector is an expression vector.
- 16. The isolated nucleic acid molecule according to claim 15 wherein the expression vector is capable of being expressed in cells derived from a eukaryote.
- 17. The isolated nucleic acid molecule according to claim 16 wherein the expression vector is further capable of being expressed in cells derived from a mammal.
- 18. The isolated nucleic acid molecule according to claim 17 wherein the expression vector is further capable of being expressed in CHO cells.

- 19. A recombinant mammalian α -N-acetylglucosaminidase or fragment or derivative thereof.
- 20. The recombinant mammalian α -N-acetylglucosaminidase according to claim 19 in substantially pure form.
- 21. The recombinant mammalian α -N-acetylglucosaminidase according to claims 19 or 20 when expressed in mammalian, yeast or insect cells.
- 22. The recombinant mammalian α-N-acetylglucosaminidase according to claim 21 when expressed in mammalian cells.
- 23. The recombinant mammalian α -N-acetylglucosaminidase according to claims 21 or 22, wherein the cells are capable of glycosylating said recombinant mammalian α -N-acetylglucosaminidase.
- 24. The recombinant mammalian α -N-acetylglucosaminidase according to claim 23 wherein the cells are capable of N-glycosylating said recombinant mammalian α -N-acetylglucosaminidase.
- 25. The recombinant mammalian α-N-acetylglucosaminidase according to claim 24 wherein the cells are CHO cells.
- 26. The recombinant mammalian α -N-acetylglucosaminidase according to any one of claims 19 to 25 wherein said recombinant α -N-acetylglucosaminidase is in a glycosylated form.
- 27. The recombinant mammalian α-N-acetylglucosaminidase according to claim 26 wherein the molecular weight of the glycosylated form as determined using SDS/PAGE is at least approximately 79 kDa.

- 28. The recombinant α-N-acetylglucosaminidase wherein the molecular weight of the glycosylated form as determined using SDS/PAGE is at least approximately 79 kDa to 89 kDa.
- 29. The recombinant mammalian α -N-acetylglucosaminidase according to any one of claims 19 to 28 comprising a sequence of amino acids substantially the same as a human α -N-acetylglucosaminidase.
- 30. The recombinant mammalian n-N-acetylglucosaminidase according to any one of claims 19 to 29 when fused to another proteinaceous molecule.
- 31. The recombinant mammalian α -N-acetylglucosaminidase according to claim 30 wherein the other proteinaceous molecule is an enzyme, reporter molecule, purification site and/or a signal sequence.
- 32. The recombinant mammalian α -N-acetylglucosaminidase according to any one of claims 19 to 31 comprising an amino acid sequence substantially as set forth in SEQ ID NO:2 or having at least 40% similarity to all or part thereof.
- 33. The recombinant mammalian α -N-acetylglucosaminidase according to claim 32 wherein the percentage similarity to SEQ ID NO:2 is at least 60%.
- 34. The recombinant mammalian α -N-acetylglucosaminidase according to claim 33 wherein the percentage similarity to SEQ ID NO:2 is at least 80%.
- 35. A recombinant α-N-acetylglucosaminidase when produced by expression of a nucleic acid molecule according to any one of claims 14 to 18.
- 36. The recombinant α -N-acetylglucosaminidase according to claim 35 when glycosylated.

- 37. A method of diagnosing a mutation in a gene which encodes α-N-acetylglucosaminidase in a human patient said method comprising contacting genomic DNA or RNA derived from said patient with one or more isolated DNA molecules or oligonucleotides comprising at least 10 contiguous nucleotides derived from SEQ ID No:1 or SEQ ID No:3 or a complementary strand thereof for a time and under conditions sufficient for hybridisation to occur and then detecting said hybridisation using a detection means.
- 38. The method according to claim 37 wherein the detection means is a reporter molecule covalently attached to the isolated DNA molecule or oligonucleotide.
- 39. The method according to claim 38 wherein the reporter molecule is ³²P, ³⁵S or biotin.
- 40. The method according to claim 37 wherein the detection means is a polymerase chain reaction format.
- The method according to claim 40 wherein the polymerase chain reaction format is selected from the list comprising SSCP, AMD, AFLP, IRS-PCR, iPCR or RT-PCR, amongst others.
- 42. The method according to claim 41 wherein the polymerase chain reaction format is SSCP.
- 43. The method according to any one of claims 37 to 42 wherein the isolated DNA molecule or oligonucleotide comprises at least 20 contiguous nucleotides derived from SEQ ID No:1 or SEQ ID No:3 or a complementary strand thereof.
- 44. The method according to claim 43 wherein the isolated DNA molecule or oligonucleotide comprises at least 50 contiguous nucleotides derived from SEQ ID No:1 or SEQ ID No:3 or a complementary strand thereof.

- 45. The method according to claim 44 wherein the isolated DNA molecule or oligonucleotide comprises at least 100 contiguous nucleotides derived from SEQ ID No:1 or SEQ ID No:3 or a complementary strand thereof.
- 46. A method for treating a patient suffering from α -N-acetylglucosaminidase deficiency said method comprising administering to said patient an effective amount of recombinant mammalian α -N-acetylglucosaminidase or an active fragment or derivative thereof.
- 47. The method according to claim 46 wherein the mammalian α -N-acetylglucosaminidase comprises a sequence of amino acids substantially the same as the amino acid sequence of human α -N-acetylglucosaminidase.
- 48. The method according to claim 47 wherein the patient is suffering from mucopolysaccharidosis type IIIB.
- 49. The method according to any one of claims 46 to 48 wherein the recombinant α -N-acetylglucosaminidase is produced in mammalian cells.
- 50. The method according to claim 49 wherein the mammalian cells are capable of glycosylating the recombinant α -N-acetylglucosaminidase produced therein.
- 51. The method according to claim 50 wherein the recombinant α -N-acetylglucosaminidase is in a glycosylated form.
- 52. The method according to claim 51 wherein the glycosylated form of the recombinant α-N-acetylglucosaminidase has a molecular weight as determined using SDS/PAGE of at least approximately 79kDa.
- 53. The method according to claim 52 wherein the glycosylated form of the

recombinant α-N-acetylglucosaminidase has a molecular weight as determined using SDS/PAGE of approximately 79 kDa to 89 kDa.

- 54. The method according to anyone of claims 46 to 53 wherein the recombinant α -N-acetylglucosaminidase comprises a sequence of amino acids substantially as set forth in SEQ ID No:2 or having at least 40% similarity to all or a part thereof.
- 55. The method according to claim 54 wherein the percentage similarity to SEQ ID NO:2 is at least 60%.
- 56. The method according to claim 55 wherein the percentage similarity to SEQ ID NO:2 is at least 80%.
- 57. A method for treating a patient suffering from α -N-acetylglucosaminidase deficiency said method comprising administering to said patient an effective amount of recombinant mammalian α -N-acetylglucosaminidase or an active fragment or derivative thereof, wherein said recombinant mammalian α -N-acetylglucosaminidase is produced by expression of a nucleic acid molecule according to any one of claims 14 to 18.
- The method according to any one of claims 46 to 57 wherein administration of the recombinant mammalian α -N-acetylglucosaminidase is by oral, intravenous, suppository, intraperetoneal, intramuscular, intranasal, intradermal or subcutaneous administration by infusion or implantation or by enzyme replacement therapy or by gene therapy.
- 59. The method according to claim 58 wherein the method of administration is by enzyme replacement therapy.
- 60. A pharmaceutical composition comprising a recombinant mammalian α-N-acetylglucosaminidase or an active fragment or derivative thereof and one or more pharmaceutically acceptable carriers and/or diluents.

- 61. The pharmaceutical composition according to claim 60 wherein the recombinant mammalian α -N-acetylglucosaminidase comprises a sequence of amino acids substantially the same as human α -N-acetylglucosaminidase.
- 62. The pharmaceutical composition according to claims 60 or 61 wherein the recombinant mammalian α -N-acetylglucosaminidase is produced in a mammalian cell.
- 63. The pharmaceutical composition according to claim 62 wherein the mammalian cell is a CHO cell line which is capable of glycosylating the recombinant mammalian α -N-acetylglucosaminidase.
- 64. The pharmaceutical composition according to any one of claims 60 to 63 wherein the α -N-acetylglucosaminidase is glycosylated.
- 65. The pharmaceutical composition according to claim 64 wherein the recombinant α-N-acetylglucosaminidase has a molecular weight as determined using SDS/PAGE of at least approximately 79 kDa.
- 66. The pharmaceutical composition according to claim 65 wherein the recombinant α-N-acetylglucosaminidase has a molecular weight as determined using SDS/PAGE of approximately 79kDa to 89 kDa.
- 67. The pharmaceutical composition according to any one of claims 60 to 66 wherein the recombinant α-N-acetylglucosaminidase comprises a sequence of amino acids substantially as set forth in SEQ ID No:2 or having at least 40% similarity to all or part thereof.
- 68. The pharmaceutical composition according to claim 67 wherein the percentage similarity to SEQ ID NO:2 is at least 60%.
- 69. The pharmaceutical composition according to claim 68 wherein the percentage

similarity to SEQ ID NO:2 is at least 80%.

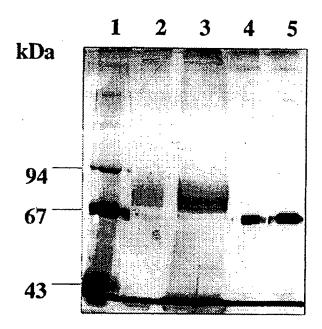
- 70. A pharmaceutical composition comprising recombinant mammalian α -N-acetylglucosaminidase or an active fragment or derivative thereof and one or more pharmaceutically acceptable carriers and/or diluents wherein said recombinant mammalian α -N-acetylglucosaminidase is produced by expression of a nucleic acid molecule according to any one of claims 14 to 18.
- 71. A pharmaceutical composition comprising recombinant mammalian α -N-acetylglucosaminidase or an active fragment or derivative thereof and one or more pharmaceutically acceptable carriers and/or diluents when used in the method according to any one of claims 46 to 59.
- Use of recombinant mammalian α -N-acetylglucosaminidase or an active fragment or derivative thereof in the manufacture of a medicament for the treatment of α -N-acetylglucosaminidase deficiency in a patient.
- 73. The use according to claim 72 wherein the recombinant mammalian α -N-acetylglucosaminidase comprises a sequence of amino acids substantially the same as the amino acid sequence of human α -N-acetylglucosaminidase.
- 74. The use according to claims 72 or 73 wherein the patient is suffering from mucopolysaccharidosis type IIIB.
- 75. The use according to claim 74 wherein the recombinant α -N-acetylglucosaminidase is expressed in mammalian cells.
- 76. The use according to claim 61 wherein the cells are CHO cells.
- 77. The use according to any one of claims 72 to 76 wherein the α -N-acetylglucosaminidase is glycosylated.

- 78. The use according to claim 77 wherein the recombinant α -N-acetylglucosaminidase has a molecular weight as determined using SDS/PAGE of at least approximately 79kDa.
- 79. The use according to claim 78 wherein the recombinant α-N-acetylglucosaminidase has a molecular weight as determined using SDS/PAGE of approximately 79 kDa to 89 kDa.
- 80. The use according to anyone of claims 72 to 79 wherein the recombinant of α -N-acetylglucosaminidase comprises a sequence of amino acids substantially as set forth in SEQ ID No:2 or has at least 40% similarity to all or a part thereof.
- 81. The use according to claim 80 wherein the percentage similarity to SEQ ID NO:2 is at least 60%.
- 82. The use according to claim 80 wherein the percentage similarity to SEQ ID NO:2 is at least 80%.
- 83. A nucleic acid molecule comprising a sequence of nucleotides encoding or complementary to a sequence encoding a polypeptide capable of hydrolysing the terminal α-N-acetylglucosaminidase residues present at the non-reducing terminus of fragments of heparan sulphate and heparin and wherein said nucleotide sequence is capable of hybridising under at least low stringency conditions to the nucleotide sequence set forth in SEQ ID NO:1.
- 84. A nucleic acid molecule comprising a sequence of nucleotides encoding or complementary to a sequence encoding a polypeptide capable of hydrolysing the terminal α-N-acetylglucosaminidase residues present at the non-reducing terminus of fragments of heparan sulphate and heparin and wherein said nucleotide sequence is capable of hybridising under at least low stringency conditions to the nucleotide sequence set forth in SEQ ID NO: 3.

- A recombinant polypeptide comprising a sequence of amino acids corresponding to the amino sequence set forth in SEQ ID NO:2 or having at least 40% similarity thereto and encoded by a nucleic acid molecule which is capable of hybridising to the nucleotide sequence set forth in SEQ ID NO:1 or SEQ ID NO:3 under at least low stringency conditions.
- 86. A genetic construct comprising the nucleic acid molecule according to any one of claims 1 to 18 or claims 83 to 84, operably connected in the sense orientation to a promoter sequence such that said genetic construct is capable of being expressed in a eukaryotic or prokaryotic cell to produce a recombinant mammalian α -N-acetylglucosaminidase or a fragment or derivative thereof.
- 87. The genetic construct according to claim 86 wherein the promoter is capable of regulating expression of the recombinant α -N-acetylglucosaminidase in a mammalian cell.
- 88. The genetic construct according to claim 87 wherein the promoter is the CMV promoter sequence or a promoter derived therefrom.
- 89. The genetic construct according to any one of claims 86 to 88 further comprising a transcription terminator sequence.
- 90. The genetic construct according to any one of claims 86 to 89 when used to express or over-express α -N-acetylglucosaminidase in a eukaryotic or prokaryotic cell.
- 91. An antibody to α -N-acetylglucosaminidase or a recombinant α -N-acetylglucosaminidase according to any one of claims 19 to 36 or an antigenic fragment thereof.
- 92. The antibody according to claim 91 further defined as a polyclonal antibody molecule.

93. The antibody according to claim 92 further defined as a monoclonal antibody molecule.

Fig. 1



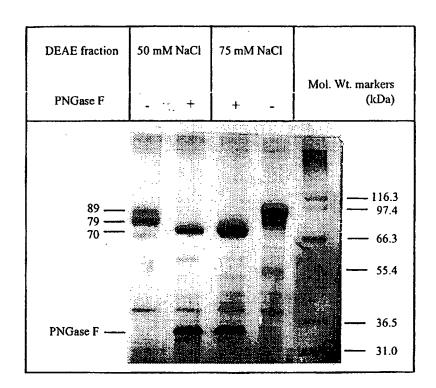


FIGURE 2

INTERNATIONAL SEARCH REPORT

International Application No.
PCT/AU 96/00747

A.	A. CLASSIFICATION OF SUBJECT MATTER						
Int Cl ⁶ : C12N 15/56, 9/24; C12Q 1/68; C07K 16/40 A61K 38/47							
According to International Patent Classification (IPC) or to both national classification and IPC							
	FIELDS SEARCHED						
Minimum documentation searched (classification system followed by classification symbols) 1C6: C12N 15/-: Keywords as below							
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Medline: see keywords below							
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) (see attached)							
C.	DOCUMENTS CONSIDERED TO BE RELEVAN	Т					
Category*	Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.				
P,X	Proc. Nat. Acad. Sci. Vol. 93, pages 6101-6105 (June 1996) H. G. Zhao et al "The Molecular Basis of Sanfilippo Syndrome Type B" (see whole document)		1-45, 83-90				
P,X	Human Molecular Genetics, Vol. 5, No. 6 pages "cloning and expression of the gene involved in (mucopolysacc-haridosis 111B)" (see whole document)	1-93					
x	Further documents are listed in the continuation of Box C	See patent family annex					
"A" docum not cor "E" earlier interna "L" docum or whit anothe "O" docum exhibit "P" docum	ent defining the general state of the art which is a sidered to be of particular relevance document but published on or after the ational filing date ent which may throw doubts on priority claim(s) ch is cited to establish the publication date of a citation or other special reason (as specified) ent referring to an oral disclosure, use, tion or other means ent published prior to the international filing at later than the priority date claimed	priority date and not in conflict with understand the principle or theory un document of particular relevance; the be considered novel or cannot be coninventive step when the document is document of particular relevance; the be considered to involve an inventive combined with one or more other suc combination being obvious to a perso	the application but cited to derlying the invention claimed invention cannot sidered to involve an taken alone claimed invention cannot step when the document is h documents, such a skilled in the art				
Date of the actual completion of the international search Date of mailing of the international search report							
10 February 1997		17 FEB 1997					
AUSTRALIAN PO BOX 200 WODEN ACT		Authorized officer KAREN AYERS					
AUSTRALIA	Facsimile No.: (06) 285 3929	Telephone No : (06) 283 2082					

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/AU 96/00747

C (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT							
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.					
<u>X</u> Y	American Journal of Genetics (1994) Volume 55, A252 (Abstract 1473) H.G. Zhao et al: "Sanfilippo Syndrome Type B cDNA and gene encoding human α-N-acetylglucosaminidase" (see whole document)	1-45, 83-90 46-82					
<u>X</u> Y	American Journal of Genetics (1995) Volume 57, A185 (Abstract 1059) H.G. Zhao et al "The Gene encoding \alpha-N-acetylglucosaminidase and mutations underlying Sanfilippo B Syndrome" (see whole document)	1-45, 83-90 46-82					
Y	Z. Klin. Chem. Klin. Biochem. Volume 13, pages 285-9 (1975), K. Von Figura et al "Serum α-N-acetylglucosaminidase: Determination, Characterization, and Corrective Activity in Sanfilippo B fibroblasts". (see whole document)	46-82					
X	Eur. J. Biochem. (1976), Volume 61, Number 2, pages 581-8, K. Von Figura and H. Kresse "Sanfilippo disease type B: presence of material cross-reacting with antibodies against alpha-N-acetylglucosaminidase" (see whole document)	91-9:					
<u>X</u> Y	J. Biochem. (1991) Volume 110, pages 842-846, T. Sasaki <u>et al</u> "A punification and partial characterisation of u-N-acetylglucosaminidase from human liver". (see whole document)	<u>91-93</u> 1-90					
		·.					
••							
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INTERNATIONAL SEARCH REPORT

International Application No.

PCT/AU 96/00747

Box B Electronic data base continued

STN:

nucleotide and amino acid sequence search

WPAT/USPM/JAPIO:

SS1 acetylglucosaminidase or acetylglucosaminohydrolase and

SS2: C12N 15/1C or San Filippo or mucopolysaccharos:

CAS online:

SS1: as above

SS2: 3/CC or 3/SX or San Filippo or mucopolysaccharos:

medline:

SS1: as above

SS2: San Filippo or mucopolysaccharos: